

MAY 7 0 2003

-1-

(Translation)

Mailed: April 15, 2003

NOTIFICATION OF REASONS FOR REJECTION

RECEIVED

Patent Application No.: 2000-247729

MAY 22 2003

Examiner's Notice Date: April 8, 2003

TECH CENTER 1600/2900

Examiner: Keiko Nagai

This application is rejected on the grounds stated below. Any opinion about the rejection must be filed within 60 DAYS of the mailing date hereof.

REASON

The invention is unpatentable under Section 29 (2) of the Patent Law, as being such that the invention could easily have been made by a person with ordinary skill in the art to which the invention pertains, on the basis of the invention described in the following publication(s) distributed in Japan or a foreign country prior to this application.

REMARKS

(1) Claim 1 : Reference 1

Reference 1 discloses a nucleotide sequence of an estrogen receptor gene of medaka fish. Here, it is easily achievable for a person having ordinary skill in the art to prepare a probe based on the nucleotide sequence of the gene discussed in Reference 1, thus obtaining polynucleotide including upstream and downstream regions of the estrogen receptor gene of medaka fish and determining its nucleotide sequence.

(2) Claim 2 : Reference 1

Reference 1 discloses a nucleotide sequence of an estrogen receptor gene of medaka fish. Here, it is easily achievable for a person having ordinary

discussed in Reference 1, thus obtaining an estrogen receptor gene of medaka fish and determining its nucleotide sequence.

(3) Claim 3 : Reference 1

Reference 1 discloses a nucleotide sequence of an estrogen receptor gene of medaka fish, and an amino acid sequence of the estrogen receptor. Here, it is easily achievable for a person having ordinary skill in the art to prepare a probe based on the nucleotide sequence of the gene discussed in Reference 1, thus obtaining an estrogen receptor gene of medaka fish and determining its nucleotide sequence and an amino acid sequence encoded by it.

(4) Claim 4 : Reference 1

As described in remarks (1) and (2) above, it is easily achievable for a person having ordinary skill in the art to obtain polynucleotide recited in Claims 1 and 2. Assembly of a recombinant vector is merely a well-known means.

(5) Claim 5 : Reference 1

Introduction of a gene to a host in order to examine an in-vivo function of a product of an isolated gene is well-known means.

The claims not mentioned in this Official Action are not rejected. If a new reason for rejection is noticed, a further Official Action will be issued.

Reference Cited:

1. Jpn. Pat. Appln. KOKAI Publication 2000-201688

Prior Art Search Report

Searched Field: IPC 7th ed. C12N 15/00

SwissProt/PIR/GeneSeq

Genbank/EMBL/DDBJ/GeneSeq

BIOSIS

MEDLINE

WPIDS

Prior-Art Document(s):

Winn R., Marine Environmental Research, vol. 46 (1-5), p.130 (1998)

pp. 192-199 (1994)

Gray M.A. et al., Environmental Toxicology and Chemistry, vol. 18(11), pp. 2587-2594 (1999)

The result of this prior art search does not constitute the reasons for rejection.

Mailing Date: April 15, 2003

整理番号 A 0 0 0 0 0 3 8 8 5

発送番号 1 2 1 2 6 4 ↓

発送日 平成 15 年 4 月 15 日

1 / 3

拒絶理由通知書

特許出願の番号	特願 2 0 0 0 - 2 4 7 7 2 9
起案日	平成 15 年 4 月 8 日
特許庁審査官	長井 啓子 9 1 2 3 4 N 0 0
特許出願人代理人	鈴江 武彦 (外 5 名) 様
適用条文	第 29 条第 2 項 15. 6. 14

この出願は、次の理由によって拒絶をすべきものである。これについて意見があれば、この通知書の発送の日から 60 日以内に意見書を提出して下さい。

理 由

この出願の下記の請求項に係る発明は、その出願前日本国内又は外国において頒布された下記の刊行物に記載された発明に基いて、その出願前にその発明の属する技術の分野における通常の知識を有する者が容易に発明をすることができたものであるから、特許法第 29 条第 2 項の規定により特許を受けることができない。

記 (引用文献等については引用文献等一覧参照)

(1) 請求項 1 : 引用文献 1

引用文献 1 には、メダカのエストロゲンレセプター遺伝子の塩基配列が開示されている。引用文献 1 記載の遺伝子の塩基配列を基にしてプローブを作成して、メダカのエストロゲンレセプター遺伝子の上流及び下流の領域を含むポリヌクレオチドを得てその塩基配列を決定することは、当業者が容易になし得る程度のことにすぎない。

(2) 請求項 2 : 引用文献 1

引用文献 1 には、メダカのエストロゲンレセプター遺伝子の塩基配列が開示されている。引用文献 1 記載の遺伝子の塩基配列を基にしてプローブを作成して、メダカのエストロゲンレセプター遺伝子を得てその塩基配列を決定することは、当業者が容易になし得る程度のことにすぎない。

(3) 請求項 3 : 引用文献 1

引用文献 1 には、メダカのエストロゲンレセプター遺伝子の塩基酸配列及び当該エストロゲンレセプターのアミノ酸配列が開示されている。引用文献 1 記載の

六島 大輔
長

発送番号 121264
発送日 平成15年 4月15日 2 / 3

遺伝子の塩基配列を基にしてプローブを作成して、メダカのエストロゲンレセプター遺伝子を得てその塩基配列及びそれがコードするアミノ酸配列を決定することは、当業者が容易になし得る程度のことにはすぎない。

(4) 請求項4：引用文献1

請求項1及び請求項2記載のポリヌクレオチドを得ることが、引用文献1の記載に基づいて当業者が容易になし得たことは、上記(1)及び(2)で説明したことおりである。組み換えベクターを構築することは常套手段にはすぎない。

(5) 請求項5：引用文献1

単離した遺伝子の産物の生体内機能を探る等の目的で、宿主に遺伝子導入することは、常套手段である。

この拒絶理由通知書中で指摘した請求項以外の請求項に係る発明については、現時点では、拒絶の理由を発見しない。拒絶の理由が新たに発見された場合には拒絶の理由が通知される。

引 用 文 献 等 一 覧

1. 特開2000-201688号公報

この拒絶理由通知書に不明な点がある場合、または、この案件について面接を希望する場合は、

特許審査第三部生命工学 長井 啓子

Tel. 03-3581-1101 (特許庁代表)

Fax. 03-3501-0491

までご連絡下さい。

先行技術文献調査結果の記録

・調査した分野 IPC第7版 C12N 15/00

SwissProt/PIR/GeneSeq

Genbank/EMBL/DDBJ/GeneSeq

MEDLINE

WPI-IDS

ID AAA92174 standard; DNA; 1728 BP.

XX
AC AAA92174; XP-002181484

XX
DT 05-JAN-2001 (first entry)

XX
DE Oryzias lapites oestrogen receptor encoding DNA SEQ ID NO:2.

XX
KW Oryzias lapites; oestrogen receptor; ds.

XX
OS Oryzias lapites.

XX
PN JP2000201688-A.

XX
PD 25-JUL-2000.

XX
PF 06-APR-1999; 99JP-0098787.

XX
PR 10-NOV-1998; 98JP-0319465.

XX
PA (SUMO) SUMITOMO CHEM CO LTD.

XX
DR WPI; 2000-567950/53.

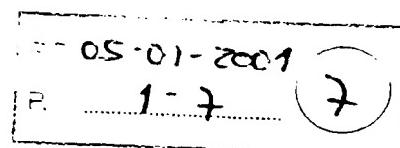
DR P-PSDB; AAB20897.

XX
PT An estrogen receptor gene and its application -

XX
PS Claim 3; Page 11-13; 23pp; Japanese.

CC
The present sequence encodes an oestrogen receptor derived from
CC Oryzias lapites. Also described are: (1) a vector comprising the
CC oestrogen receptor gene; (2) a transformant prepared by introducing
CC the oestrogen receptor gene or vector from (1) into a host cell;
CC (3) a method for the preparation of an oestrogen receptor comprising
CC culturing the transformant from (2) to produce the oestrogen receptor;
CC and (4) a method for the evaluation of oestrogen receptor-activating
CC ability of a chemical substance in which the chemical substance is
CC reacted with a transformant prepared by introducing a reporter gene
CC connected downstream of a transcription controlling region containing
CC an oestrogen response sequence and the above oestrogen receptor gene to
CC an oestrogen-nonendogenous host cell. The transformant can be used for
CC the evaluation of oestrogen receptor-activating ability of a chemical
CC substance.

XX
SQ Sequence 1728 BP; 378 A; 514 C; 497 G; 339 T; 0 other;
atgtaccctg aagagagccg gggttcttga ggggtggctg ctgtggaccc tttggaaagg 60
acgtacgact atgcggcccc caaccctgcc acgactcccc ttacagccca gtcccgacacc 120
ggctactact ctgtctccctt gaaaacaac ggacccccc cagaaggcag tctgcagttcc 180
ctggcagtg gggcagcgg ccctctggtg tttgtgcctt ccagccccag actcagtccc 240
tttatgcattt caccctggcc ccactatctg gaaaccattt ccacccccgtt ttacagatcc 300
agccaccagg gagccctccag ggaggaccag tgccgctccc gggaggacac gtgcagctg 360
ggggagtttag gcgcggagc cggggctggg gggtttgaga tggccaaaga cacgcgtttc 420
tgcccgctgt gcagcacta cgcctctggg taccactatg gggtgtggtc ttgtgagggc 480
tgcaaggctt tcttcaagag gagcatccag ggtcacaatg actatatgtg cccagcgacc 540
aatcgtgca ctattgacag aaatcgaagg aagggtgtc aggcttgcg tccttaggaag 600
tgttacgaag tggaaatgt gaaaggcggt gtgcgcagg accgcattcg catttacgg 660
cgtgacaaac ggcggacagg cggtggatgat ggagacaagg ttgttaaagggt tcaggagcat 720
aaaacgggtgc attatgtgg aaggaaacgc agcagcacag gaggaggagg aggaggagga 780
ggaggaagac tgcgtgtgac cagcataccct cctgagcagg tgctgtctt ccttcaggcc 840
gccgagcccc cgatactctg ctcgcgtcag aagttgagcc gaccgtacac cgaggtcacc 900
atgatgaccc tgcgttccatg cttgcgttccatg aaggagctgg tccacatgtat cgcctggcc 960
aagaagctcc cagggtttctt gcaagctgtcc ctgcacgtac aggtgtgtgt gctggagac 1020
tcgtggctgg aggtgtctcat gatcgccctc atttggaggtt ccattccactg tcccgaaag 1080
ctcatcttgc cacaagaccc catcctggac aggaatgagg gagactgcgtt ggaaggcatg 1140



acggagatct tcgacatgct gctggccact gcttcccgt tccgtgtgt ccaaactcaaa 1200
cctgaggaat tcgtctgcct caaagctatt attttactca actccgggtgc tttttctttc 1260
tgccaccggca ccatggagcc acttcacaac agcgcgccggg ttcagagcat gctggacacc 1320
atcacagacg cactcattca ttacatcagt cagtcgggtt acttggccca ggagcaggcg 1380
agacggcagg cccagccgct cctgctgctc tccccacateca ggcacatgag caacaaaggc 1440
atggagcacc tctacagcat gaagtgcagg aacaaagtcc ctctttatga ctcctactg 1500
gagatgtctg atgeccaccg cctgeaccac cccgtcagag cccccccatgc cttgtcccaa 1560
gtcgacagag accctccctc caccaggcgc ggcgggggtg gaatcgctcc cggttctata 1620
tcagcatctc gaggcagaat cgagagtcgg agcagagggc cctttgctcc cagtgtcctt 1680
cagtatggag ggtcgctcc tgactgcacc cccggcccttc aagactga 1728

//

>>GSN:AAA92174 Oryzias lapites oestrogen receptor (1728 nt)
initn: 8586 init1: 8586 opt: 8586 Z-score: 8271.0 bits: 1544.2 E(): 0
99.653% identity (99.653% ungapped) in 1728 nt overlap (211-1938:1-1728)

190 200 210 220 230 240
EP0111 CGCCTCTCGCCCCGTGACCCCTCGGTGACATGTACCCCTGAAGAGAGCCGGGTTCTGGA
:::
GSN:AA ATGTACCCCTGAAGAGAGCCGGGTTCTGGA

10 20 30

250 260 270 280 290 300
EP0111 GGGGTGGCTGCTGTGGACTTTTGGAAAGGGACGTACGACTATGCCGCCCCAACCTGCC
:::
GSN:AA GGGGTGGCTGCTGTGGACCTTTGGAAAGGGACGTACGACTATGCCGCCCCAACCTGCC
40 50 60 70 80 90

310 320 330 340 350 360
EP0111 ACGACTCCCTTTACAGCCAGTCCAGCACCGGCTACTACTCTGCCTCCCTGGAAACAAAC
:::
GSN:AA ACGACTCCCTTTACAGCCAGTCCAGCACCGGCTACTACTCTGCCTCCCTGGAAACAAAC
100 110 120 130 140 150

370 380 390 400 410 420
EP0111 GGACCCCCCTCAGAAGGCAGTCTGCAGTCCCTGGGAGTGGGCCACGAGCCCTCTGGTG
:::
GSN:AA GGACCCCCCTCAGAAGGCAGTCTGCAGTCCCTGGGAGTGGGCCACGAGCCCTCTGGTG
160 170 180 190 200 210

430 440 450 460 470 480
EP0111 TTTGTGCCCTCCAGCCCCAGACTCAGTCCCTTATGCATCCACCCAGCCACCACTATCTG
:::
GSN:AA TTTGTGCCCTCCAGCCCCAGACTCAGTCCCTTATGCATCCACCCAGCCACCACTATCTG
220 230 240 250 260 270

490 500 510 520 530 540
EP0111 GAAACCACCTCCACGCCGTTACAGATCCAGCACCCAGGGAGCCTCCAGGGAGGACCAAG
:::
GSN:AA GAAACCACCTCCACGCCGTTACAGATCCAGCACCCAGGGAGCCTCCAGGGAGGACCAAG
280 290 300 310 320 330

550 560 570 580 590 600
EP0111 TCGGGCTCCCGGGAGGACACGTGCAGCCTGGGGAGTTAGGCGCCGGAGCCGGGGCTGGG
:::
GSN:AA TCGGGCTCCCGGGAGGACACGTGCAGCCTGGGGAGTTAGGCGCCGGAGCCGGGGCTGGG
340 350 360 370 380 390

610 620 630 640 650 660
EP0111 GGGTTTGAGATGGCAAAGACACCGCGTTCTGCGCCGTGTCAGCGACTACGCCTCTGGG
:::
GSN:AA GGGTTTGAGATGGCAAAGACACCGCGTTCTGCGCCGTGTCAGCGACTACGCCTCTGGG
400 410 420 430 440 450

670 680 690 700 710 720
EP0111 TACCACTATGGGGTGTGGTCTTGTGAGGGCTGCAAGGCCTTCTCAAGAGGAGCATCCAG
:::
GSN:AA TACCACTATGGGGTGTGGTCTTGTGAGGGCTGCAAGGCCTTCTCAAGAGGAGCATCCAG
460 470 480 490 500 510

730 740 750 760 770 780
EP0111 GGTCAACAATGACTATATGTGCCAGCGACCAATCAGTGCACATTGACAGAAATCGGAGG
:::
GSN:AA GGTCAACAATGACTATATGTGCCAGCGACCAATCAGTGCACATTGACAGAAATCGAAGG
520 530 540 550 560 570

	790	800	810	820	830	840
EP0111	AAGAGCTGCCAGGCTTGTCTTACGAAGTGGAAATGATGAAAGGC	GGT				
	580	590	600	610	620	630
	850	860	870	880	890	900
EP0111	GTGCGCAAGGACCGCATTGCTTCAGGCGTGACAAACGGCGACAGGC	GTTGGT	GAT			
	640	650	660	670	680	690
	910	920	930	940	950	960
EP0111	GGAGACAAGGTTGTAAAGGGTCAGGAGCATAAAACGGTCATTATG	ATGGAAGGAAACGC				
	700	710	720	730	740	750
	970	980	990	1000	1010	1020
EP0111	AGCAGCACAGGAGGAGGAGGAGGAGGAGGAGGAGGAGGAGGAGG	AAGACTGTC	TGTGACCAC	GATACCT		
	760	770	780	790	800	810
	1030	1040	1050	1060	1070	1080
EP0111	CCTGAGCAGGTGCTGCTCCCTCPTCAGGGCGCCGAGCCCCCGA	FACTCTGCTCGC	GTCAG			
	820	830	840	850	860	870
	1090	1100	1110	1120	1130	1140
EP0111	AAGTTGAGCCGACCGTACACCGAGGTCACCATGATGACCC	CTGCTCACCA	GATGGCAGAC			
	880	890	900	910	920	930
	1150	1160	1170	1180	1190	1200
EP0111	AAGGAGCTGGTCCACATGATCGCCTGGCCAAGAACGCTCCCAGG	TTCTGCAGCTG	CC			
	940	950	960	970	980	990
	1210	1220	1230	1240	1250	1260
EP0111	CTGCACGATCAGGTGCTGCTGCTGGAGAGCTCGTGG	CTGGACCTGCTCATG	ATGATECCCCCTC			
	1000	1010	1020	1030	1040	1050
	1270	1280	1290	1300	1310	1320
EP0111	ATTTGGAGGTCCATCCACTGTCEGGCAAGCTCATCTTG	GAACAAGAC	CTCATCCTGGAC			
	1060	1070	1080	1090	1100	1110
	1330	1340	1350	1360	1370	1380
EP0111	AGGAATGAGGGAGACTGCGTGGAAAGGCATGACGGAGA	ACTCTGCACATGCTG	GGCCACT			
	1120	1130	1140	1150	1160	1170
	1390	1400	1410	1420	1430	1440
EP0111	GCTTCCCGCTTCCGTGTGCTCAAAC	TCATAACCTGAGGA	ATTCTGCTG	CCTCAAGCTATT		
	1180	1190	1200	1210	1220	1230

	1450	1460	1470	1480	1490	1500
EP0111	ATTTTACTCAACTCCGGTGCTTTCTTCTGCACCGGCACCATGGAGCCACTTCACAAAC
GSN:AA	ATTTTACTCAACTCCGGTGCTTTCTGGACCGGCACCATGGAGCCACTTCACAAAC	1240	1250	1260	1270	1280
	1510	1520	1530	1540	1550	1560
EP0111	AGCGCGGCCGGTTCAGAGCATGGACACCATCACAGAGGAACTCATTACATCAGT
GSN:AA	AGCGCGGCCGGTTCAGAGCATGGACACCATCACAGACGCACTCATTACATCAGT	1300	1310	1320	1330	1340
	1570	1580	1590	1600	1610	1620
EP0111	CAGTCGGGTTACTTGGCCCAGGAGCAGGCAGACGGCAGGCCAGCTGCTCCTGCTGCTC
GSN:AA	CAGTCGGGTTACTTGGCCCAGGAGCAGGCAGGCCAGCCGCTCTGCTGCTC	1360	1370	1380	1390	1400
	1630	1640	1650	1660	1670	1680
EP0111	TCCCACATCAGGCACATGAGCAACAAAGGCATGGAGAACCTTAAGGATGAAGTCCAG
GSN:AA	TCCCACATCAGGCACATGAGCAACAAAGGCATGGAGAACCTTAAGGATGAAGTCCAG	1420	1430	1440	1450	1460
	1690	1700	1710	1720	1730	1740
EP0111	AACAAAGTC CCTCTTTATGACCTCTACTGAGATGCTCGATGCCAACCGCCTGCAAC
GSN:AA	AACAAAGTC CCTCTTTATGACCTCTACTGAGATGCTCGATGCCAACCGCCTGCAAC	1480	1490	1500	1510	1520
	1750	1760	1770	1780	1790	1800
EP0111	CCCGTCAGAGCACCCAGTCCTTCTCCAGTCAGAGACCTCTECACCAGGAG
GSN:AA	CCCGTCAGAGCACCCAGTCCTTGTCCAAAGTCGACAGAGACCTCTCCACCAGCAGC	1540	1550	1560	1570	1580
	1810	1820	1830	1840	1850	1860
EP0111	GGCGGGGGTGAATCGCTCCGGPTCTATATCAGCATCTCGAGGCAGAATCGAGACTCG
GSN:AA	GGCGGGGGTGAATCGCTCCGGTTCTATATCAGCATCTCGAGGCAGAATCGAGACTCG	1600	1610	1620	1630	1640
	1870	1880	1890	1900	1910	1920
EP0111	AGCAGAGGCCCTTGCTCCAGTGTCTTCAGTATGGAGGGTCCGCTCTGACTGCACC
GSN:AA	AGCAGAGGCCCTTGCTCCAGTATGGAGGGTCCGCTCTGACTGCACC	1650	1670	1680	1690	1700
	1930	1940	1950	1960	1970	1980
EP0111	CCGGCCCTCAAGACTGAGCACACAGTCCAAGGCCCTTTTGTGGCTCAAGGGTTCAAG
GSN:AA	CCGGCCCTCAAGACTGA	1720				

14

ID AAB20897 standard; Protein; 575 AA.
XX
AC AAB20897;
XX
DT 05-JAN-2001 (first entry)
XX
DE Oryzias lapites oestrogen receptor protein SEQ ID NO:1.
XX
KW Oryzias lapites; oestrogen receptor.
XX
OS Oryzias lapites.
XX
PN JP2000201688-A.
XX
PD 25-JUL-2000.
XX
PF 06-APR-1999; 99JP-0098787.
XX
PR 10-NOV-1998; 98JP-0319465.
XX
PA (SUMO) SUMITOMO CHEM CO LTD.
XX
DR WPI; 2000-567950/53.
DR N-PSDB; AAA92174.
XX
PT An estrogen receptor gene and its application -
XX
PS Claim 1; Page 9-10; 23pp; Japanese.
XX
CC The present sequence represents an oestrogen receptor derived from
CC Oryzias lapites. Also described are: (1) a vector comprising the
CC oestrogen receptor gene; (2) a transformant prepared by introducing
CC the oestrogen receptor gene or vector from (1) into a host cell;
CC (3) a method for the preparation of an oestrogen receptor comprising
CC culturing the transformant from (2) to produce the oestrogen receptor;
CC and (4) a method for the evaluation of oestrogen receptor-activating
CC ability of a chemical substance in which the chemical substance is
CC reacted with a transformant prepared by introducing a reporter gene
CC connected downstream of a transcription controlling region containing
CC an oestrogen response sequence and the above oestrogen receptor gene to
CC an oestrogen-nonendogenous host cell. The transformant can be used for
CC the evaluation of oestrogen receptor-activating ability of a chemical
CC substance.
XX
SQ Sequence 575 AA;
SQ 37 A; 38 R; 10 N; 27 D; 0 B; 18 C; 24 Q; 29 E; 0 Z; 58 G; 19 H;
SQ 22 I; 61 L; 24 K; 19 M; 14 F; 32 P; 58 S; 29 T; 4 W; 20 Y; 27 V;
SQ 0 Others;
SQ mypeesrgsg gvaavdlleg tydyaapnpa ttplysqsst gyysapletn gppsegslqs
lqsgptspolv fpvssprlsp fmhppshhyl ettstpvrys shqgasredq cgsredtcs1
gelgagagag gfemakdtrf cavcsdyasg yhygvwsceg ckaffkrsiq ghndymcpat
nqctidmrnr kgcqacrlrk cyeygrmkqg vrkdiririlr rdkrxtqvqd qdkvvkqgeh
ktvhhydgrkr sstggggggg ggrlsvtsip peqvllllqg aepplcsrc klsrppteve
mmtlltsmad kelvhmiawa kklpgflqls lhdqvllles swlevlmigl iwrsihcpkg
lifaqidild rnegdcvegm teifdmllat asrfrvlklk peefvclkai illnsgafsf
ctgtmeplnh saavqsmldt itdalihyis qsgylaqeqa rrqaqp111 shirhmsnk
mehlysmkck nkvpolyd111 emldahrlhh pvrarpqslsq vdrdppstss ggggiapgsi
sasrgriesp srqpfapsvl qyggsrpdc palqd

11

>>GSP:AAB20897 Oryzias lapites oestrogen receptor (575 aa)
initn: 3905 init1: 3905 opt: 3905 Z-score: 2879.0 bits: 544.9 E(): 2.1e-152
Smith-Waterman score: 3905; 99.478% identity (99.478% ungapped) in 575 aa overlap
(210-1934:1-575)

210	240	270	300	330	360
EP0111	MYPEESRGSGVAAVDLLEGTYDYAAPNPATTPLYSQSSTGYYSAPLETNGPPSEGLQS				
GSP:AA	MYPEESRGSGVAAVDLLEGTYDYAAPNPATTPLYSQSSTGYYSAPLETNGPPSEGLQS				
	10	20	30	40	50
					60
390	420	450	480	510	540
EP0111	LGGSGPTSPPLVFVPSSPRLSPEMHPPSHYLETTSTPVYRSSHQGASREDQCGSREDTCSL				
GSP:AA	LGGSGPTSPPLVFVPSSPRLSPEMHPPSHYLETTSTPVYRSSHQGASREDQCGSREDTCSL				
	70	80	90	100	110
					120
570	600	630	660	690	720
EP0111	GELGAGAGAGGFEMAKDTRFCAVCSDYASGYHYGVWSCEGCKAFFKRSIQGHNDYMCPAT				
GSP:AA	GELGAGAGAGGFEMAKDTRFCAVCSDYASGYHYGVWSCEGCKAFFKRSIQGHNDYMCPAT				
	130	140	150	160	170
					180
750	780	810	840	870	900
EP0111	NQCTIDRNRRKSCQACRLRKCYEVGMMKGVRKDRIRILRRDKRRTGVGDGDKVVKQEH				
GSP:AA	NQCTIDRNRRKGQCQACRLRKCYEVGMMKGVRKDRIRILRRDKRRTGVGDGDKVVKQEH				
	190	200	210	220	230
					240
930	960	990	1020	1050	1080
EP0111	KTVHYDGRKRSSTGGGGGGGGRLSVTSIPPEQVLLLLQGAEPPILCSRQKLSRPYTEVT				
GSP:AA	KTVHYDGRKRSSTGGGGGGGGRLSVTSIPPEQVLLLLQGAEPPILCSRQKLSRPYTEVT				
	250	260	270	280	290
					300
1110	1140	1170	1200	1230	1260
EP0111	MMTLLTSMADKELVHMIWAKKLPGLQLSLHDQVLLLESSWLEVLMIGLIWRSIHCPGK				
GSP:AA	MMTLLTSMADKELVHMIWAKKLPGLQLSLHDQVLLLESSWLEVLMIGLIWRSIHCPGK				
	310	320	330	340	350
					360
1290	1320	1350	1380	1410	1440
EP0111	LIFAQDLILDRNEGDCVEGMTEIFDMLLATASRFRVLKLKPEEFVCLKAIILLNSGAFSF				
GSP:AA	LIFAQDLILDRNEGDCVEGMTEIFDMLLATASRFRVLKLKPEEFVCLKAIILLNSGAFSF				
	370	380	390	400	410
					420
1470	1500	1530	1560	1590	1620
EP0111	CTGTMEPLHNSAAVQSMLEDTITDALIHYISQSGYLAQEQRQAQPLLLLShIRHMSNKG				
GSP:AA	CTGTMEPLHNSAAVQSMLEDTITDALIHYISQSGYLAQEQRQAQPLLLLShIRHMSNKG				
	430	440	450	460	470
					480
1650	1680	1710	1740	1770	1800
EP0111	MEHLYSMKCKNKVPLYDLLLEM DALHRLHHPVRA PQSLSQVDRDP PSTSSGGGIAPGS				
GSP:AA	MEHLYSMKCKNKVPLYDLLLEM DALHRLHHPVRA PQSLSQVDRDP PSTSSGGGIAPGS				
	490	500	510	520	530
					540
1830	1860	1890	1920		
EP0111	SASRGRIESPSRGPFAPS V LQYGGSRPD CTPAL QD				
GSP:AA	SASRGRIESPSRGPFAPS V LQYGGSRPD CTPAL QD				
	550	560	570		

ID AAA92175 standard; DNA; 1863 BP.
XX
AC AAA92175; XP-002181483

P.D.QS-07-2001
P. 1-9- 9

10

DT 05-JAN-2001 (first entry) --

XX DE Oryzias lapites oestrogen receptor encoding DNA SEQ-ID NO:4.

XX KW Oryzias lapites; oestrogen receptor ds.

XX OS Oryzias lapites.

XX PN JP2000201688-A.

XX PD 25-JUL-2000.

XX PF 06-APR-1999; 99JP-0098787.

XX PR 10-NOV-1998; 98JP-0319465.

XX PA (SUMO) SUMITOMO CHEM CO LTD.

XX DR WPI; 2000-567950/53.

DR P-PSDB; AAB20898.

XX PT An estrogen receptor gene and its application

XX PS Claim 4; Page 15-17; 23pp; Japanese.

XX CC The present sequence encodes an oestrogen receptor derived from Oryzias lapites. Also described are: (1) a vector comprising the oestrogen receptor gene; (2) a transformant prepared by introducing the oestrogen receptor gene or vector from (1) into a host cell; (3) a method for the preparation of an oestrogen receptor comprising culturing the transformant from (2) to produce the oestrogen receptor; and (4) a method for the evaluation of oestrogen receptor-activating ability of a chemical substance in which the chemical substance is reacted with a transformant prepared by introducing a reporter gene connected downstream of a transcription controlling region containing an oestrogen response sequence and the above oestrogen receptor gene to an oestrogen-nonendogenous host cell. The transformant can be used for the evaluation of oestrogen receptor-activating ability of a chemical substance.

XX SQ Sequence 1863 BP; 406 A; 565 C; 531 G; 361 T; 0 other;

atgagtaaga	gacagagctc	ggtcgcagatc	aggcagactgt	tccggaccgc	actcagatcc	60
aggatcagcc	cagccctcc	agagctggag	accctctccc	cacctcgct	ctcgccccgt	120
gacc	ccctcg	gigacatgt	ccctgaagag	agccggggtt	ctggagggtt	180
gac	tttgg	aagggacgta	cgactatgcc	gcccccaacc	ctgcccacgac	240
agcc	atcca	gcacccggcta	ctactctgct	ccccctggaaa	caaacggacc	300
ggc	actgc	agtccctggg	cagtggccg	acgagccctc	tggtgtttgt	360
ccc	ccatca	gtcccttat	qcatccaccc	agccaccaact	atctggaaac	420
ccg	tttaca	gatccagcc	ccagggagcc	tccagggagg	accagtgcgg	480
gac	cttgca	gcctggggqa	gttagggcgc	ggagccgggg	ctgggggtt	540
aa	agacacgc	gtttctgcgc	cgtgtgcagc	gactacgcct	ctatgggtg	600
tt	tttgc	agggctgcaa	ggccttcttc	aagaggagca	tccagggtca	660
gg	ccat	atgtgcccag	cgaccaatca	gtgcactatt	gacagaaatc	720
gg	tttgc	ggaagtgtta	cgaagtggga	atgatgaaag	gaagaagg	780
gg	tttgc	atcgcat	tacggcgtga	caaacggcgg	ctgggtgtcg	840
gg	tttgc	atcgcat	acaggcgttg	gtgatggaga	caagggttga	900
gg	tttgc	aagggtcagg	agcataaaac	ggtgcattat	aatggaaagga	960
gg	tttgc	ggaggaggag	ggaggaggag	aagactgtct	gtgaccagaca	1020
gg	tttgc	ctcccttc	aggcgccga	gcccccgata	tttctgcgc	1080
gg	tttgc	tacaccatgt	gaccctgctc	accagcatgg	gacacaagga	1140
gg	tttgc	atgatgcct	ggccaagaa	gctcccaagg	gtccctgca	

ctgctgctgg agagctcgta	gctggaggtg ctcatgatcg	gcctcatttg gaggtccatc	1200
cactgtccc ggaagctcat	cttgcacaa gacctcatcc	tggacaggaa tgagggagac	1260
tgcgttggaaag	gcatgacgga gatcttcgac	atgctgctgg ccactgcttc	1320
gtgctcaaacc	tcaaacctga ggaattcgta	tgccctcaaag ctattattt	1380
ggtgttttt	cttctgcac cggcaccatg	gagccacttc acaacagcgc	1440
agcatgctgg	acaccatcac agacgcactc	attcattaca tcagtcagtc	1500
gcccaggagc	aggcgagacg gcaggcccag	ccgctcctgc tgctctccca	1560
atgagcaaca	aaggcatgga gcacctctac	agcatgaagt gcaagaacaa	1620
tatgacctcc	tactggagat gctcgatgcc	caccgcctgc accacccgt	1680
cagtccttgt	cccaagtcga cagagaccct	ccctccacca gcagcggcgg	1740
gctcccggtt	ctatatcagc atctcgaggc	agaategaga gtccgagcag	1800
gctcccggtt	tccttcagta tggagggtcg	gcaccccggc cttcaagac	1860
tga			1863

//

>>GSN:AAA92175 Oryzias lapites oestrogen receptor (1863 nt)
initn: 9261 initl: 9261 opt: 9261 Z-score: 8922.0 bits: 1664.7 E(): 0
99.678% identity (99.678% ungapped) in 1863 nt overlap (76-1938:1-1863)

50 60 70 80 90 100
EP0111 CGTGTGCGCAGCACATCTG**A**GGATGATTCA**T**GTAAGAGACAGAGCTCGGTGCAGATC
:::
GSN:AA ATGAGTAAGAGACAGAGCTCGGTGCAGATC
10 20 30

110 120 130 140 150 160
EP0111 AGGCAGCTGTTCGGACCAGCACTCAGATCCAGGA**T**CAGGCCAGCCTCCTCAAGAGCTGGAG
:::
GSN:AA AGGCAGCTGTTCGGACCAGCACTCAGATCCAGGA**T**CAGGCCAGCCTCCTCAAGAGCTGGAG
40 50 60 70 80 90

170 180 190 200 210 220
EP0111 ACCCTCTCCCCACCTCGCCTCTCGCCCCGTGACCCCCCTCGGTGACATGTACCCCTGAAGAG
:::
GSN:AA ACCCTCTCCCCACCTCGCCTCTCGCCCCGTGACCCCCCTCGGTGACATGTACCCCTGAAGAG
100 110 120 130 140 150

230 240 250 260 270 280
EP0111 AGCCGGGGTTCTGGAGGGGTGGCTGCTGTGACTTTTGGAAGGGACGTACGACTATGCC
:::
GSN:AA AGCCGGGGTTCTGGAGGGGTGGCTGCTGTGACCTTTTGGAAGGGACGTACGACTATGCC
160 170 180 190 200 210

290 300 310 320 330 340
EP0111 GCCCCCACCCCTGCCACGACTCCCCTTAACGCCAGTCAGCAGCCGCTACTACTCTGCT
:::
GSN:AA GCCCCCACCCCTGCCACGACTCCCCTTAACGCCAGTCAGCAGCCGCTACTACTCTGCT
220 230 240 250 260 270

350 360 370 380 390 400
EP0111 CCCCTGGAAACAAACGGACCCCCCTCAGAAGGCAGTCTGCAGTCCTGGCAGTGGCCG
:::
GSN:AA CCCCTGGAAACAAACGGACCCCCCTCAGAAGGCAGTCTGCAGTCCTGGCAGTGGCCG
280 290 300 310 320 330

410 420 430 440 450 460
EP0111 ACGAGCCCTCTGGTGTGTTGTGCCCTCCAGCCCCAGACTCAGTCCTTATGCATCCACCC
:::
GSN:AA ACGAGCCCTCTGGTGTGTTGTGCCCTCCAGCCCCAGACTCAGTCCTTATGCATCCACCC
340 350 360 370 380 390

470 480 490 500 510 520
EP0111 AGCCACCACTATCTGGAAACCACCTCCACGCCGTTACAGATCCAGCCACCAGGGAGCC
:::
GSN:AA AGCCACCACTATCTGGAAACCACCTCCACGCCGTTACAGATCCAGCCACCAGGGAGCC
400 410 420 430 440 450

530 540 550 560 570 580
EP0111 TCCAGGGAGGACCA**G**TCGGCTCCGGAGGACACGTGCAGCCTGGGGAGTTAGGCC
:::
GSN:AA TCCAGGGAGGACCA**G**TCGGCTCCGGAGGACACGTGCAGCCTGGGGAGTTAGGCC
460 470 480 490 500 510

590 600 610 620 630 640
EP0111 GGAGCCGGGCTGGGGTTGAGATGGCAAAGACACGCCGTTCTGCCCGTGTGCAGC
:::
GSN:AA GGAGCCGGGCTGGGGTTGAGATGGCAAAGACACGCCGTTCTGCCCGTGTGCAGC
520 530 540 550 560 570

650 660 670 680 690 700

EP0111 GACTACGCCCTGGGTACCACTATGGGTGAGGGCTGCAAGGCCTCTTC
 :::::::::::::::::::::
 GSN:AA GACTACGCCCTGGGTACCACTATGGGTGAGGGCTGCAAGGCCTCTTC
 580 590 600 610 620 630
 710 720 730 740 750 760
 EP0111 AAGAGGAGCATCCAGGGTACAATGACTATATGTGCCAGCGACCAATCAGTGCCTATT
 :::::::::::::::::::::
 GSN:AA AAGAGGAGCATCCAGGGTACAATGACTATATGTGCCAGCGACCAATCAGTGCCTATT
 640 650 660 670 680 690
 770 780 790 800 810 820
 EP0111 GACAGAAATCGGAGGAAGAGCTGCCAGGCTTGTGCTTAGGAAGTGTTACGAAGTGGGA
 :::::::::::::::::::::
 GSN:AA GACAGAAATCGAAGGAAGGGCTGTCAGGCTTGTGCTTAGGAAGTGTTACGAAGTGGGA
 700 710 720 730 740 750
 830 840 850 860 870 880
 EP0111 ATGATGAAAGGCGGTGTGCGCAAGGACCGCATTGCACTTACGGCGTACAAACGGCGG
 :::::::::::::::::::::
 GSN:AA ATGATGAAAGGCGGTGTGCGCAAGGACCGCATTGCACTTACGGCGTACAAACGGCGG
 760 770 780 790 800 810
 890 900 910 920 930 940
 EP0111 ACAGGCCTGGTGTGGAGACAAGGTTGTAAGGGTCAGGAGCATAAAACGGTGCATTAT
 :::::::::::::::::::::
 GSN:AA ACAGGCCTGGTGTGGAGACAAGGTTGTAAGGGTCAGGAGCATAAAACGGTGCATTAT
 820 830 840 850 860 870
 950 960 970 980 990 1000
 EP0111 GATGGAAGGAAACGCAGCACAGGAGGAGGAGGAGGAGGAGGAGGAGGAAAGACTGTCT
 :::::::::::::::::::::
 GSN:AA GATGGAAGGAAACGCAGCACAGGAGGAGGAGGAGGAGGAGGAGGAGGAGGAAAGACTGTCT
 880 890 900 910 920 930
 1010 1020 1030 1040 1050 1060
 EP0111 GTGACCAGCATACCTCCTGAGCAGGTGCTGCTCCTCCTTCAGGGCGCCGAGCCCCCGATA
 :::::::::::::::::::::
 GSN:AA GTGACCAGCATACCTCCTGAGCAGGTGCTGCTCCTCCTTCAGGGCGCCGAGCCCCCGATA
 940 950 960 970 980 990
 1070 1080 1090 1100 1110 1120
 EP0111 CTCTGCTCGCGTCAGAAGTTGAGCCACCGTACACCGAGGTACCATGATGACCTGCTC
 :::::::::::::::::::::
 GSN:AA CTCTGCTCGCGTCAGAAGTTGAGCCACCGTACACCGAGGTACCATGATGACCTGCTC
 1000 1010 1020 1030 1040 1050
 1130 1140 1150 1160 1170 1180
 EP0111 ACCAGCATGGCAGACAAGGAGCTGGTCCACATGATGCCCTGGCCAAGAAGCTCCCAGGT
 :::::::::::::::::::::
 GSN:AA ACCAGCATGGCAGACAAGGAGCTGGTCCACATGATGCCCTGGCCAAGAAGCTCCCAGGT
 1060 1070 1080 1090 1100 1110
 1190 1200 1210 1220 1230 1240
 EP0111 TTTCTGCAGCTGTCCCTGCACGATCAGGTGCTGCTGGAGAGCTCGTGGCTGGAGGTG
 :::::::::::::::::::::
 GSN:AA TTTCTGCAGCTGTCCCTGCACGATCAGGTGCTGCTGGAGAGCTCGTGGCTGGAGGTG
 1120 1130 1140 1150 1160 1170
 1250 1260 1270 1280 1290 1300
 EP0111 CTCATGATCGGCCTCATTGGAGGTCCATCCACTGTCCCAGGAAGCTCATCTTGACCAA
 :::::::::::::::::::::
 GSN:AA CTCATGATCGGCCTCATTGGAGGTCCATCCACTGTCCCAGGAAGCTCATCTTGACCAA
 1180 1190 1200 1210 1220 1230

1310	1320	1330	1340	1350	1360
EP0111 GACCTCATCCTGGACAGGAATGAGGGAGACTGCGTGGAAAGGCATGACGGAGATCTCGAC					
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
GSN:AA GACCTCATCCTGGACAGGAATGAGGGAGACTGCGTGGAAAGGCATGACGGAGATCTCGAC					
1240	1250	1260	1270	1280	1290
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
1370	1380	1390	1400	1410	1420
EP0111 ATGCTGCTGCCACTGCTTCCCGTTCCGTGTCAAACCTCAAACCTGAGGAATTGTC					
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
GSN:AA ATGCTGCTGCCACTGCTTCCCGTTCCGTGTCAAACCTCAAACCTGAGGAATTGTC					
1300	1310	1320	1330	1340	1350
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
1430	1440	1450	1460	1470	1480
EP0111 TGCCTCAAAGCTATTATTTACTCAACTCCGGTGCTTTCTTCTGCACCGGCACCATG					
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
GSN:AA TGCCTCAAAGCTATTATTTACTCAACTCCGGTGCTTTCTTCTGCACCGGCACCATG					
1360	1370	1380	1390	1400	1410
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
1490	1500	1510	1520	1530	1540
EP0111 GAGCCACTTCACAACAGCGCGGGTTCAGAGCATGCTGGACACCATCACAGACGCACTC					
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
GSN:AA GAGCCACTTCACAACAGCGCGGGTTCAGAGCATGCTGGACACCATCACAGACGCACTC					
1420	1430	1440	1450	1460	1470
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
1550	1560	1570	1580	1590	1600
EP0111 ATTCAATTACATCAGTCAGTCGGTTACTTGGCCCAGGAGCAGGGAGACGGCAGGCCAG					
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
GSN:AA ATTCAATTACATCAGTCAGTCGGTTACTTGGCCCAGGAGCAGGGAGACGGCAGGCCAG					
1480	1490	1500	1510	1520	1530
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
1610	1620	1630	1640	1650	1660
EP0111 CTGCTCCTGCTCTCCACATCAGGCACATGAGCAACAAAGGCATGGAGCACCTCTAC					
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
GSN:AA CCGCTCCTGCTCTCCACATCAGGCACATGAGCAACAAAGGCATGGAGCACCTCTAC					
1540	1550	1560	1570	1580	1590
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
1670	1680	1690	1700	1710	1720
EP0111 AGCATGAAGTGCAGAACAAAGTCCCTCTTATGACCTCCTACTGGAGATGCTCGATGCC					
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
GSN:AA AGCATGAAGTGCAGAACAAAGTCCCTCTTATGACCTCCTACTGGAGATGCTCGATGCC					
1600	1610	1620	1630	1640	1650
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
1730	1740	1750	1760	1770	1780
EP0111 CACCGCCTGCACCACCCCGTCAGAGCACCCAGTCCTGTCCCAGTCACAGAGACCT					
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
GSN:AA CACCGCCTGCACCACCCCGTCAGAGCCCCAGTCCTGTCCCAGTCACAGAGACCT					
1660	1670	1680	1690	1700	1710
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
1790	1800	1810	1820	1830	1840
EP0111 CCCTCCACCAGCAGCGGGGGGTGAATCGCTCCCGTTCTATATCAGCATCTCGAGGC					
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
GSN:AA CCCTCCACCAGCAGCGGGGGGTGAATCGCTCCCGTTCTATATCAGCATCTCGAGGC					
1720	1730	1740	1750	1760	1770
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
1850	1860	1870	1880	1890	1900
EP0111 AGAATCGAGAGTCGAGCAGAGGCCCTTGCTCCAGTGTCTTCAGTATGGAGGGTCG					
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
GSN:AA AGAATCGAGAGTCGAGCAGAGGCCCTTGCTCCAGTGTCTTCAGTATGGAGGGTCG					
1780	1790	1800	1810	1820	1830
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
1910	1920	1930	1940	1950	1960
EP0111 CGTCCTGACTGCACCCGGCCCTCAAGACTGAGCACACAGTCCAAGGCCCTTTTGT					
::: :::::::::::::::::::: :::::::::::::::::::: ::::::::::::::::::::					
GSN:AA CGTCCTGACTGCACCCGGCCCTCAAGACTGA					
1840	1850	1860			

1970 1980 1990 2000 2010 2020
EP0111 GGCTCAAGGGTTCAGGTTGGGACAAGGTGATGCTTGATTAAATTTAAGAATTATTTATA

ID AAB20898 standard; Protein; 620 AA.
XX
AC AAB20898;
XX
DT 05-JAN-2001 (first entry)
XX
DE Oryzias lapites oestrogen receptor protein SEQ ID NO:3.
XX
KW Oryzias lapites; oestrogen receptor.
XX
OS Oryzias lapites.
XX
PN JP2000201688-A.
XX
PD 25-JUL-2000.
XX
PF 06-APR-1999; 99JP-0098787.
XX
PR 10-NOV-1998; 98JP-0319465.
XX
PA (SUMO) SUMITOMO CHEM CO LTD.
XX
DR WPI; 2000-567950/53.
DR N-PSDB; AAA92175.
XX
PT An estrogen receptor gene and its application
XX
PS Claim 2; Page 13-15; 23pp; Japanese.
XX
CC The present sequence represents an oestrogen receptor derived from
CC Oryzias lapites. Also described are: (1) a vector comprising the
CC oestrogen receptor gene; (2) a transformant prepared by introducing
CC the oestrogen receptor gene or vector from (1) into a host cell;
CC (3) a method for the preparation of an oestrogen receptor comprising
CC culturing the transformant from (2) to produce the oestrogen receptor;
CC and (4) a method for the evaluation of oestrogen receptor-activating
CC ability of a chemical substance in which the chemical substance is
CC reacted with a transformant prepared by introducing a reporter gene
CC connected downstream of a transcription controlling region containing
CC an oestrogen response sequence and the above oestrogen receptor gene to
CC an oestrogen-nonendogenous host cell. The transformant can be used for
CC the evaluation of oestrogen receptor-activating ability of a chemical
CC substance.
XX
SQ Sequence 620 AA:
SQ 39 A; 44 R; 10 N; 29 D; 0 B; 18 C; 27 Q; 31 E; 0 Z; 60 G; 19 H;
SQ 24 I; 67 L; 25 K; 20 M; 15 F; 43 P; 67 S; 30 T; 4 W; 20 Y; 28 V;
SQ 0 Others:
mskrqssvqi rqlfgpalrs rispassele tlspprlspr dplgdmypee srgsgggvaav
dllegtydya apnpattply sqsstgyysa pletngppse gslqslgsqo tsplvfvpss
prlspfmhpp shhylettst pvyrsshsgga sredqcgssre dtcslgelga gagaggfema
kdtrfcavcs dyasgyhygv ~~wsccegckaff~~ krsiqghndy mcpatnacti dmrrkgcqa
crlrkcyevg mmkggvrkdr iirilrrdkrr tgvgdgdkvv kgqehktvhq dgrkrssstgg
gggggggrls vtsippeqvl lllogeoppi lceqohqsp ytevtmtll tsmadkelvh
miawakklpg flqlslhdqv lllesswlev lmigliwrsi hcpgklifaq dlildmnedg
cvegmtieifd mlatasrfr viklkpeefv cikaiiilns gefsfctgtm epihnsaavq
smlldtitdal ihyisqsgyl aqeqarrqaq plllshirh msnkgmehly smkcknkvp
ydllemlda hrlhhpvrap qslsqvdrdp pstssgggi apgsisasrg riespsrgpf
apsvlqyqgs rpdcptpalqd

//

1 Yell
variation of seq ID 1
VS
Seq ID 3 of 'SP'... 20168

>>GSP:AAB20898 Oryzias lapites oestrogen receptor (620 aa)
initn: 4198 initl: 4198 opt: 4198 Z-score: 3093.7 bits: 584.7 E(): 2.3e-164
Smith-Waterman score: 4198: 99.516% identity (99.516% ungapped) in 620 aa overlap
(75-1934:1-620)

90	120	150	180	210	240
EP0111 MSKRQSSVQIRQLFGPALRSRISPASSELETLSPPRLSPRDPLGDMYPEESRGSGGVAAV					
GSP:AA MSKRQSSVQIRQLFGPALRSRISPASSELETLSPPRLSPRDPLGDMYPEESRGSGGVAAV					
10	20	30	40	50	60
270	300	330	360	390	420
EP0111 DFLEGTYDYAAPNPATTPLYSQSSTGYYSALENGPPSEGSLQSLGSGPTSPLVFVPSS					
GSP:AA DLLEGTYDYAAPNPATTPLYSQSSTGYYSALENGPPSEGSLQSLGSGPTSPLVFVPSS					
70	80	90	100	110	120
450	480	510	540	570	600
EP0111 PRLSPFMHPPSHYLETSTPVYRSSHQGASREDQCGSREDTCSLGEAGAGAGGFEMA					
GSP:AA PRLSPFMHPPSHYLETSTPVYRSSHQGASREDQCGSREDTCSLGEAGAGAGGFEMA					
130	140	150	160	170	180
630	660	690	720	750	780
EP0111 KDTRFCAVCSDYASGYHYGVWSCEGCKAFFKRSIQQGHNDYMCPATNQCTIDRNRRKSCQA					
GSP:AA KDTRFCAVCSDYASGYHYGVWSCEGCKAFFKRSIQQGHNDYMCPATNQCTIDRNRRKGCQA					
190	200	210	220	230	240
810	840	870	900	930	960
EP0111 CRLRKCYEVGMMKGVRKDRIRILRRDKRTGVGDGDKVVKGQEHTVHYDGRKRSSTGG					
GSP:AA CRLRKCYEVGMMKGVRKDRIRILRRDKRTGVGDGDKVVKGQEHTVHYDGRKRSSTGG					
250	260	270	280	290	300
990	1020	1050	1080	1110	1140
EP0111 GGGGGGGRLSVTSIPPEQVLLLQGAEPPILCRSRQKLSRPYTEVTMMTLLTSMADKELVH					
GSP:AA GGGGGGGRLSVTSIPPEQVLLLQGAEPPILCRSRQKLSRPYTEVTMMTLLTSMADKELVH					
310	320	330	340	350	360
1170	1200	1230	1260	1290	1320
EP0111 MIAWAKKLPGFLQLSLHDQVLLLESSWLEVLMIGLIWRSIHCPGKLIFAQDLILDRNEGD					
GSP:AA MIAWAKKLPGFLQLSLHDQVLLLESSWLEVLMIGLIWRSIHCPGKLIFAQDLILDRNEGD					
370	380	390	400	410	420
1350	1380	1410	1440	1470	1500
EP0111 CVEGMTEIFDMILLATASRFRVLKLKEEFVCLKAIILLNSGAFSFCTGTMEPLHNSAAVQ					
GSP:AA CVEGMTEIFDMILLATASRFRVLKLKEEFVCLKAIILLNSGAFSFCTGTMEPLHNSAAVQ					
430	440	450	460	470	480
1530	1560	1590	1620	1650	1680
EP0111 SMLDTITDALIHYISQSGYLAQEQRQAQPLLLLSHIRHMSNKGMELYSMKCKNVPL					
GSP:AA SMLDTITDALIHYISQSGYLAQEQRQAQPLLLLSHIRHMSNKGMELYSMKCKNVPL					
490	500	510	520	530	540
1710	1740	1770	1800	1830	1860
EP0111 YDLLEMLDAHRLHHPVRAPOQLSQVDRDPSTSSGGGIAPGSISASRGRIESPSRGPF					
GSP:AA YDLLEMLDAHRLHHPVRAPOQLSQVDRDPSTSSGGGIAPGSISASRGRIESPSRGPF					
550	560	570	580	590	600

1890	1920
EP0111 APSVLQYGGSRPDCTPALQD	
: : : : : : : : : : : :	
GSP:AA APSVLQYGGSRPDCTPALQD	
610	620

This entry is from:

SWALL (SPTR)

General Description References Comments Links Keywords Features Sequence

24

[CSV](#)[FASTA](#)

SWISS-PROT

General information

Entry name: ESR1_ORYLA

Accession number: P50241

01-10-1996

Created: Rel. 34, 1-OCT-1996

1-5

S

Sequence update: Rel. 37, 15-DEC-1998

Annotation update: Rel. 40, 16-OCT-2001

Description and origin of the Protein

Description: ESTROGEN RECEPTOR (ER) (ESTRADIOL RECEPTOR) (ER-ALPHA).

Gene name(s): ESR OR NR3A1 OR MER.

Organism source: Oryzias latipes (Medaka fish).

Taxonomy: Eukaryota; Metazoa; Chordata; Craniata; Vertebrata; Euteleostomi; Actinopterygii; Teleostei; Euteleostei; Neoteleostei; Acanthomorpha; Acanthoperciformes; Atherinomorpha; Beloniformes; Adrianichthyidae; Oryziinae;

NCBI TaxID: 8090

References

[1] Okada,H., Kawahara,T., Yamashita,I., RL Submitted (APR-1998) to the EMBL/GenBank/DDBJ databases.

Position: RP SEQUENCE FROM N.A.

Comments: RC STRAIN=D-RR; TISSUE=LIVER;

[2] Kawahara,T., Yamashita,I., RT "Oryzias latipes genomic DNA for estrogen receptor.RL Submitted to the EMBL/GenBank/DDBJ databases.

Position: RP SEQUENCE FROM N.A.

Comments

FUNCTION	THE STEROID HORMONES AND THEIR RECEPTORS ARE INVOLVED IN THE REGULATION OF EUKARYOTIC GENE EXPRESSION AND A CELLULAR PROLIFERATION AND DIFFER IN TARGET TISSUES.
----------	--

SUBUNIT	BINDS DNA AS A HOMODIMER. CAN FORM HETERO-DIMER WITH ER-BETA (BY SIMILARITY).
---------	---

SUBCELLULAR LOCATION	NUCLEAR.
----------------------	----------

DOMAIN	COMPOSED OF THREE DOMAINS: A MODIFIED N-TERMINAL DOMAIN, A DNA-BINDING DOMAIN AND A C-TERMINAL STEROID-BINDING DOMAIN.
--------	--

SIMILARITY	BELONGS TO THE NUCLEAR HORMONE RECEPTOR FAMILY. NR3 SUBFAMILY.
------------	--

Copyright

This SWISS-PROT entry is copyright. It is produced through a collaboration between the Swiss Bioinformatics and the EMBL outstation - the European Bioinformatics Institute. There are no restrictions on its use by non-profit institutions as long as its content is in no way modified and this statement is present.

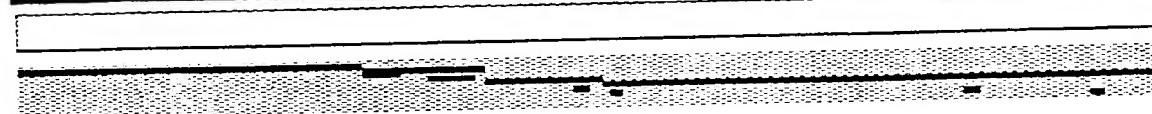
Database cross-references

EMBL	D28954;BAA25900.1;-.
HSSP	AB033491;BAA86925.1;-.
InterPro	P03372;1HCP.
	IPR000536;Hormone_rec_lig.
	IPR001292;Oest_recep.
	IPR001723;Strdhormone_receptor.
	IPR001628;zf-C4.
	PF00104;hormone_rec;1.
Plant	PF02159;Oest_recep;1.
	PF00105;zf-C4;1.
PRINTS	PR00398;STRDHORMONER.
	PR00047;STROIDFINGER.
SMART	SM00430;HOLI;1.
	SM00399;ZnF_C4;1.
PROSITE	PS00031;NUCLEAR_RECECTOR;1.

Keywords

Receptor, Transcription regulation; DNA-binding; Nuclear protein; Zinc-finger; Steroid-binding;

Features



Key	Begin	End	Length	Description
DOMAIN	1	185	185	MODULATING.
DNA_BIND	186	251	66	NUCLEAR RECEPTOR-TYPE.
ZN_FING	186	206	21	C4-TYPE.
ZN_FING	222	246	25	C4-TYPE.
DOMAIN	252	314	63	HINGE.
DOMAIN	315	620	306	STEROID-BINDING.
DOMAIN	299	307	9	POLY-GLY.
DOMAIN	320	323	4	POLY-LEU.
DOMAIN	511	515	5	POLY-LEU.
DOMAIN	576	579	4	POLY-GLY.

Sequence information

Length: 620 aa, molecular weight: 67729 Da, CRC64 checksum: DDCBD18C2B2BA522
MSKRQSSVQI RQLFGPALRS RISPASSELE TLSPPRLSPR DPLGDMYPEE SRGSGGVA DFLEGTYDYA APNPATTPLY SQSSTGYYSA PLETNGPPSE GSLQLSGSGP TSPLVFVPS PRLSPFMHPP SHHYLETTST PVYRSSHQGA SREDQCGSRE DTCSLGEELGA GAGAGGF KDTRFCAVCS DYASGYHYGV WSCEGCKAFF KRSIQGHNDY MCPATNQCTI DRNRRKS CRLRKCYEVG MMKGGVRKDR IRILRRDKRR TGVGDGDKVV KGQEHKTVHY DGRKRSS GGGGGGGRLS VTSIPPEQLV LLLQGAEPPI LCSRQKLSRP YTEVTMMTLL TSMADKELV MIAWAKKLPG FLQLSLHDQV LLLESSWLEV LMIGLIWRSI HCPGKLIFAQ DLILDRNEGD

CVEGMTEIFD MLLATASRFR VLKLKPEEFV CLKAIILLNS GAFSFCTGTM EPLHNSAAVQ
SMLDTITDAL IHYISQSGYL AQEQARRQAQ LLLLSSHIRH MSNKGMELHY SMKCKNVPI
YDLLEMLDA HRLHHPVRAP QSLSQVDRDP PSTSSGGGGI APGSISASRG RIESPSRGF
APSVLQYGGS RPDCTPALQD

620

//

[General Description](#) [References](#) [Comments](#) [Links](#) [Keywords](#) [Features](#) [Sequence](#)

SRS 6.1.3 | feedback

>>SWALL:ESR1_ORYLA ESTROGEN RECEPTOR (ER) (ESTRAD (620 aa)
initn: 4219 init1: 4219 opt: 4219 z-score: 3109.1 bits: 587.6 E(): 3.1e-165
Smith-Waterman score: 4219; 100.000% identity (100.000% ungapped) in 620 aa
overlap (75-1934:1-620)

90	120	150	180	210	240
EP0111 MSKRQSSVQIRQLFGPALRSRISPASSELETLSPPKLSPRDPLGDMYPEESRGSGGVAAV	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::
SWALL: MSKRQSSVQIRQLFGPALRSRISPASSELETLSPPRLSPRDPLGDMYPEESRGSGGVAAV	10	20	30	40	50
					60
270	300	330	360	390	420
EP0111 DFLEGTYDYAAPNPATTPLYSQSSTGYYSAPLETNGPPSEGSLQSLGSGPTSPLVFVPSS	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::
SWALL: DFLEGTYDYAAPNPATTPLYSQSSTGYYSAPLETNGPPSEGSLQSLGSGPTSPLVFVPSS	70	80	90	100	110
					120
450	480	510	540	570	600
EP0111 PRLSPFMHPPSHHYLETTSTPVYRSSHQGASREDQCGSREDTCSLGELGAGAGAGGFEMA	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::
SWALL: PRLSPFMHPPSHHYLETTSTPVYRSSHQGASREDQCGSREDTCSLGELGAGAGAGGFEMA	130	140	150	160	170
					180
630	660	690	720	750	780
EP0111 KDTRFCAVCSDYASGYHYGVWSCEGCKAFFKRSIQGHNDYMCPATNQCTIDRNRRKSCQA	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::
SWALL: KDTRFCAVCSDYASGYHYGVWSCEGCKAFFKRSIQGHNDYMCPATNQCTIDRNRRKSCQA	190	200	210	220	230
					240
810	840	870	900	930	960
EP0111 CRLRKYEVGMMKGVRKDRIRILRRDKRRTGVGDGDKVVKGQEhkTVHYDGRKRSSTGG	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::
SWALL: CRLRKYEVGMMKGVRKDRIRILRRDKRRTGVGDGDKVVKGQEhkTVHYDGRKRSSTGG	250	260	270	280	290
					300
990	1020	1050	1080	1110	1140
EP0111 GGGGGGGRLSVTSIPPEQVLLLQGAEPPILCRQKLSRPYTEVTMMTLLTSMADKELVH	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::
SWALL: GGGGGGGRLSVTSIPPEQVLLLQGAEPPILCRQKLSRPYTEVTMMTLLTSMADKELVH	310	320	330	340	350
					360
1170	1200	1230	1260	1290	1320
EP0111 MIAWAKKLPGLQLSLHDQVLLLESSWLEVLMIGLIWRSIHCPGKLIFAQDLILDRNEGD	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::
SWALL: MIAWAKKLPGLQLSLHDQVLLLESSWLEVLMIGLIWRSIHCPGKLIFAQDLILDRNEGD	370	380	390	400	410
					420
1350	1380	1410	1440	1470	1500
EP0111 CVEGMTEIFDMLLATASRFRVLKLKPEEFVCLKAIILLNSGAFSFCTGTMEPLHNSAAVQ	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::
SWALL: CVEGMTEIFDMLLATASRFRVLKLKPEEFVCLKAIILLNSGAFSFCTGTMEPLHNSAAVQ	430	440	450	460	470
					480
1530	1560	1590	1620	1650	1680
EP0111 SMLDTITDALIHYISQSGYLAQEQRQAQLLLLLSHIRHMSNKGMELYSMKCKNVPL	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::
SWALL: SMLDTITDALIHYISQSGYLAQEQRQAQLLLLLSHIRHMSNKGMELYSMKCKNVPL	490	500	510	520	530
					540
1710	1740	1770	1800	1830	1860
EP0111 YDLLLEMLDAHRLHHPVRAPOQLSQVDRDPPSTSSGGGIAPGSISASRGRIESPSRGPF	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::	:::::::::::::::::::
SWALL: YDLLLEMLDAHRLHHPVRAPOQLSQVDRDPPSTSSGGGIAPGSISASRGRIESPSRGPF	550	560	570	580	590
					600

1890 1920
EP0111 APSVLQYGGSRPDCTPALQD
:::::::
SWALL: APSVLQYGGSRPDCTPALQD
610 620

発送番号 121264
発送日 平成15年 4月15日 3 / 3

・先行技術文献

Winn R., Marine Environmental Research, vol. 46(1-5), p. 130 (1998)

Takagi S. et al., Molecular Marine Biology and Biotechnology, vol. 3(4),
pp. 192-199 (1994)

Gray M. A. et al., Environmental Toxicology and Chemistry, vol. 18(11),
pp. 2587-2594 (1999)

この先行技術文献調査結果の記録は、拒絶理由を構成するものではない。

Aryl Hydrocarbon Receptor is Required for Prevention of Blood Clotting and for the Development of Vasculature and Bone in the Embryos of Medaka Fish, *Oryzias latipes*

Toshiyuki Kawamura and Ichiro Yamashita*

*Center for Gene Science, Hiroshima University, Kagamiyama 1-4-2,
Higashi-Hiroshima 739-8527, Japan*

ABSTRACT—The aryl hydrocarbon receptor (AHR) is a member of ligand-activated transcription factors and conserved among vertebrates. To investigate the role of AHR in fish development, medaka embryos were treated with agonist (2,3,7,8-tetrachlorodibenzo-*p*-dioxin), antagonists (α -naphthoflavone and resveratrol), and inhibitor (piperonyl butoxide) of cytochromes (Cysts) P450 encoded by a battery of target genes. These embryos were found to have similar abnormal phenotypes. Among the most consistent phenotypes were blood clotting and malformation of bone that were associated with vascular damages. These results thus indicate that control of AHR is important for proper development of fish embryos. AHR may control levels of Cysts P450 that are responsible for synthesis and metabolism of a toxic compound that caused the abnormal phenotypes. Complementary DNA fragments encoding AHR homologs were cloned from medaka embryos. AHR-specific mRNA was ubiquitously expressed in embryos and adult tissues.

Key words: aryl hydrocarbon receptor, blood clotting, bone formation, cytochrome P450, dioxin.

INTRODUCTION

Planar halogenated hydrocarbons, such as 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD), are notorious environmental pollutants that are extremely toxic to early stages of vertebrate development (Peterson *et al.*, 1993). Hallmark signs of TCDD toxicity in fish sac fry are yolk sac edema, slowed blood flow, hemorrhage, and growth retardation culminating in mortality (Cantrell *et al.*, 1996; Henry *et al.*, 1997; Hornung *et al.*, 1999). Vascular damage, as assessed by TCDD-induced apoptotic cell death, is a key physiological mediator of the embryo toxicity (Cantrell *et al.*, 1996; Cantrell *et al.*, 1998). These chemicals bind to a ligand-dependent transcriptional factor called the aryl hydrocarbon receptor (AHR), resulting in the activation of a battery of genes encoding various cytochromes (Cysts) P450 that are responsible for degradation of the environmental contaminants (Hankinson, 1995; Guiney *et al.*, 1997; Guiney *et al.*, 2000). AHR is conserved among vertebrates, thus, may have arisen in an ancestral vertebrate as a detoxification system.

Although, to date, an endogenous ligand for AHR has not been found, AHR is ubiquitously expressed in most

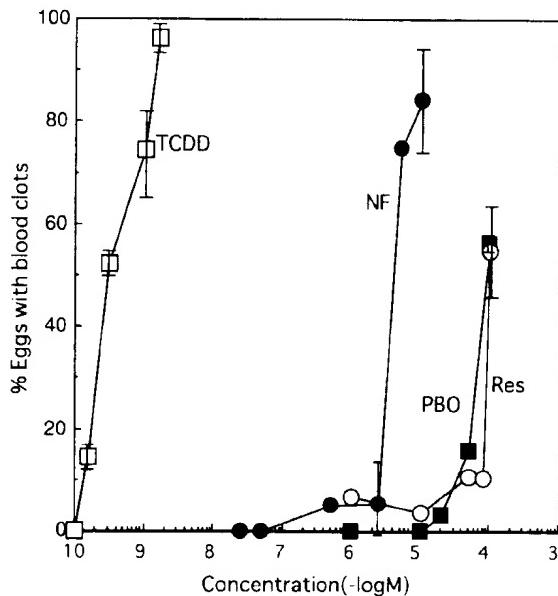
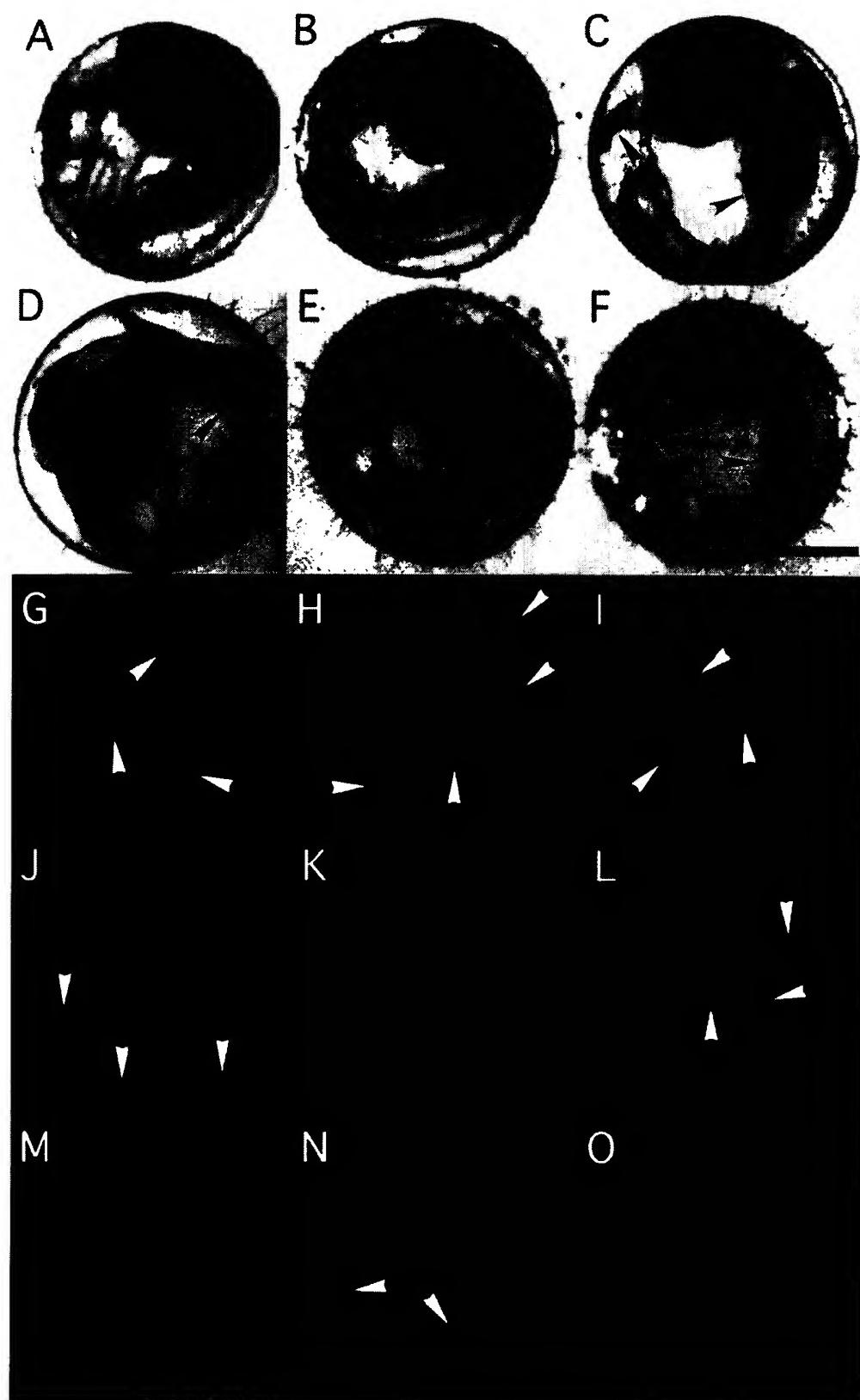


Fig. 1. Effects of 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD), α -naphthoflavone (NF), resveratrol (Res), and piperonyl butoxide (PBO) on blood clotting during the embryo stage. Eggs were treated with TCDD, NF, Res, or PBO at the indicated concentrations until 6, 6, 4, or 5 dpf, respectively, and counted for blood clots.

* Corresponding author: Tel. +81-824-24-6271;
FAX. +81-824-22-7184.
E-mail: iyama@hiroshima-u.ac.jp



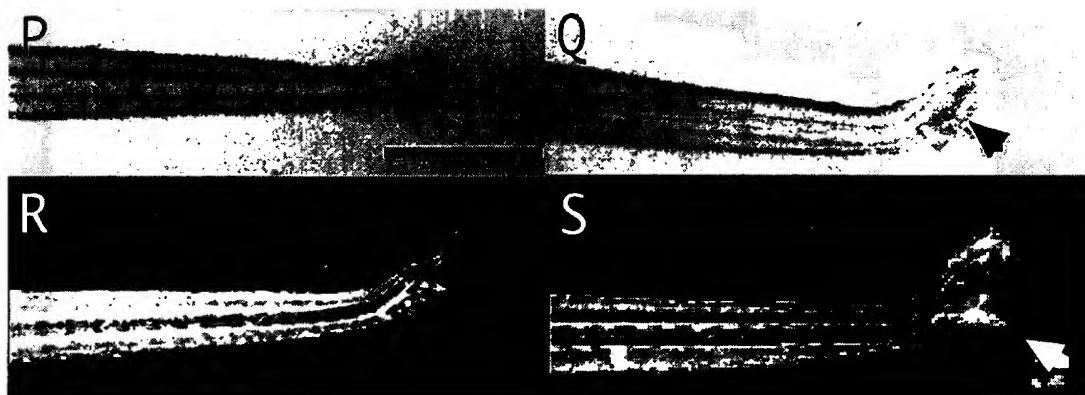


Fig. 2. Photographs of blood clots, yolk vein, and fin. Eggs and fry were treated with 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD), α -naphthoflavone (NF), resveratrol (Res), or piperonyl butoxide (PBO) as follows and photographed for blood clots (**A–F**), yolk vein under green fluorescence (**G–O**), and fin (**P–S**): (**A**) mock-treated, 5 dpf; (**B**, **C**) 1.55 nM TCDD, 5 and 7 dpf; (**D**) 10 μ M NF, 5 dpf; (**E**) 100 μ M PBO, 5 dpf; (**F**) 100 μ M Res, 4 dpf; (**G–I**) mock-treated at 3, 5, and 7 dpf; (**J**, **K**) 1.55 nM TCDD, 5 and 7 dpf; (**L**, **M**) 10 μ M NF, 3 and 5 dpf; (**N**, **O**) 100 μ M PBO, 4 and 5 dpf; (**P**, **R**) mock-treated, 5-day post-hatching; and (**Q**, **S**) 0.155 nM TCDD, 5-day post-hatching. Arrows indicate blood clots (**B–F**, and **Q**), yolk veins (**G–J**, **L**, and **N**), and the constricted fin (**S**). Bar, 0.5 mm.

organs and cells in the body (Rowlands and Gustafsson, 1997). However, there is only a limited knowledge of developmental and physiological functions of AHR in the mouse (Gonzalez and Fernandez-Salguero, 1998), although the role of AHR in detoxification of environmental aryl hydrocarbons has been extensively studied *in vitro* (Hankinson, 1995). AHR-null mice were resistant to the acute toxicity (Fernandez-Salguero *et al.*, 1996) of and the teratogenic response (Mimura *et al.*, 1997) to TCDD, and found to have a number of abnormal phenotypes such as decreased accumulation of lymphocytes in the spleen and lymph nodes and reduction in liver size that are associated with accelerated rates of apoptosis (Fernandez-Salguero *et al.*, 1995), and difficulties in reproduction (Abbott *et al.*, 1999; Robles *et al.*, 2000). Thus, AHR is involved in the toxicity of and the teratogenesis by TCDD *in vivo*, and plays an important role in the development of the liver and the immune system, and in reproduction. However, no such function has been elucidated in other vertebrates.

Here we re-evaluated the role of AHR in chemical toxicity of TCDD in medaka fish embryos because there have been no pharmacological studies in fish using antagonist and also examined for any possible developmental and physiological function of AHR in medaka fish embryos using antagonists and Cyt P450 inhibitor. We found that AHR mediates TCDD toxicity such as blood clotting, malformation of bone, and regression of blood vessels, and that AHR is required for the embryonic development of vasculature and bone. To our knowledge, this is the first report of the developmental role of AHR in lower vertebrates.

MATERIALS AND METHODS

Fish and embryo culture

We used the d-rR strain of medaka fish, *O. latipes* (Kawahara and Yamashita, 2000). The fish were maintained at 25–26°C under

artificial photo-period of 14L 10D, and fed by powdered Tetramin (Tetra). Eggs were collected within 12 hr postfertilization (hpf), rinsed with tap water, and immersed in Yamamoto's salt solution (Yamamoto, 1969) with or without test chemicals. At least 30 eggs were used in each experiment. TCDD was purchased from Cambridge Isotope Laboratories, Inc. Antagonists, α -naphthoflavone (NF) (Gasiewicz and Rucci, 1991; Merchant *et al.*, 1993) and resveratrol (Res) (Ciolino *et al.*, 1998; Casper *et al.*, 1999; Singh *et al.*, 2000), were from Sigma. Cyt P450 inhibitor, piperonyl butoxide (PBO) (Dahl and Hodgson, 1979; Testa and Jenner, 1981; Adams *et al.*, 1993), was from Tokyo Kasei Kogyo Co. These reagents were dissolved in acetone. The stock solutions were diluted over 1,000-fold with Yamamoto's solution and added to eggs of 12 hpf for NF, Res, and PBO or of 24 hpf for TCDD. The solvent was added to the mock-treated eggs as a control. The reducing agent, N-acetyl cysteine (NAC) (Sigma), was dissolved in Yamamoto's solution and added to 12 hpf eggs. Eggs and fry were cultured under the same condition as above (except without feed) and inspected for blood clotting under a dissecting microscope. Eggs and fry in which blood clots formed were counted.

Data are presented as mean \pm SEM. Statistical significance between values of control and experiment was assessed by Student's *t*-test.

Observation of blood vessels

In order to observe the development of blood vessels, eggs were fixed with 4% paraformaldehyde for 3 days and observed under green fluorescence with a filter set (excitation filter, 546/10 nm, barrier filter, 590 nm) in Leica MZ FLIII stereo-fluorescence microscope. The fixed eggs were also dechorionated with forceps and stained with hematoxylin.

Bone staining

In order to observe the bone development, calcified bone was stained with alizarin S essentially as described (Takeuchi, 1960). In brief, fish were anesthetized with 0.015% phenylurethane, skinned with forceps, treated with 2% KOH for 24 hr, and finally stained with 0.1% alizarin S solution. After washing in tap water, the fish were successively transferred to 50% and 70%, and finally embedded in 100% glycerin. Anesthetized fry were directly treated with 2% KOH for 2 h, fixed in 4% paraformaldehyde for 24 hr, then stained with alizarin S.

Isolation of cDNAs encoding medaka AHR homologs

As PAS domain of AHR is highly conserved among vertebrates (Rowlands and Gustafsson, 1997), a corresponding region of cDNA was amplified with degenerated oligonucleotides (AhR-A1 and AhR-B1) as described (Hahn and Karchner, 1995) using total RNA from 6-day postfertilization (dpf) medaka embryos. The cDNA fragment was cloned in plasmid and sequenced. Based on the sequence, nested oligonucleotides were designed and 5' and 3' RACEs (rapid amplification of cDNA ends) were performed on the same RNA by using 5' and 3' RACE Systems (GIBCO BRL), yielding the remainder of the coding sequence, 5' and 3' untranslated regions, and polyadenylation sequence.

RNA analysis

Total RNA was extracted from embryos and adult tissues as described (Kawahara *et al.*, 2000). RT-PCR (reverse transcription-polymerase chain reaction) analysis was done as described (Kawahara *et al.*, 2000) with the primers as follows for generation of the 437-bp cDNA encoding a part of PAS domain: poly(dT) oligonucleotide used for RT, and 5'-CCAGCAGGAGTCAGGAGGA and 5'-ATTTTACCCCTTGCCTCACCA for PCR. Amplified DNA was electrophoresed in 1% agarose gel and stained with ethidium bromide.

RESULTS

AHR mediates the toxic effects of TCDD on vascular development

We re-evaluated the toxic effects of TCDD on medaka embryos. To do this, embryos (1 dpf) were immersed in saline solution for medaka containing increasing concentrations of TCDD, and observed for any abnormal phenotype under a dissecting microscope (Fig. 1). Clearly visible signs of blood clotting were apparent after 4 days in caudal veins of TCDD-treated embryos (Fig. 2B), although blood cells were circulating in vasculature (Fig. 2J) but at a reduced rate. Blood clots were also found in yolk veins after 6 days (Fig. 2C), at that time, vascular structure was almost absent (Fig. 2K). In control embryos, yolk veins were apparent at 3 dpf (Fig. 2G) and developed progressively in a curve structure (Fig. 2A, H and I). Very small blood clots were occasionally found in yolk veins of normal embryos (less than 3%), but not scored in this study. These results are consistent with the previous observation that TCDD induces apoptosis of blood vessels (Cantrell *et al.*, 1996).

If TCDD induced the vascular damage through activation of AHR, the antagonist (NF) would reduce the extent to which blood clotting was detected. For this purpose, two different experiments were done, in which embryos were treated with high (1.55 nM) or medial (0.775 nM) concentration of TCDD (Fig. 3A or B, respectively). For both cases, addition of NF effectively suppressed blood clotting but only transiently (Fig. 3A and B). However, in the latter case, NF markedly enhanced the hatching success of TCDD-treated embryos, giving rise to almost complete hatching (Fig. 3C). These results indicate that TCDD-induced vascular damage is mediated through activation of AHR.

It is well known that TCDD-bound AHR activates transcription of a battery of genes encoding Cys P450. If these

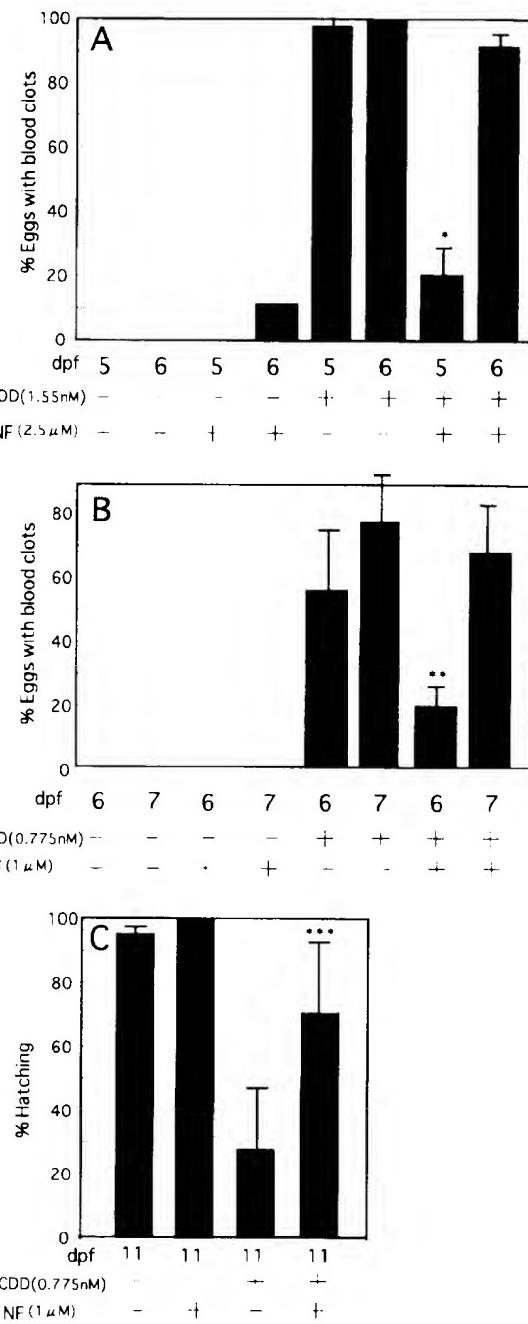


Fig. 3. Suppression by α -naphthoflavone (NF) of 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD)-induced blood clotting and mortality. **(A)** Eggs were treated with 1.55 nM TCDD and 2.5 μ M NF until 5 and 6 dpf as indicated, and examined for blood clotting. *P<0.01. **(B)** Eggs were treated with 0.775 nM TCDD and 1 μ M NF until 6 and 7 dpf as indicated, and examined for blood clotting. **P<0.2. **(C)** Eggs were treated as described in **(B)**, and examined for hatching rate at 11 dpf. ***P<0.05.

enzymes were involved in the TCDD-induced toxicity, an inhibitor of P450 would reduce the rate of TCDD-induced blood clotting. We therefore examined the ability of PBO to provide protection against high concentration (1.55 nM) of

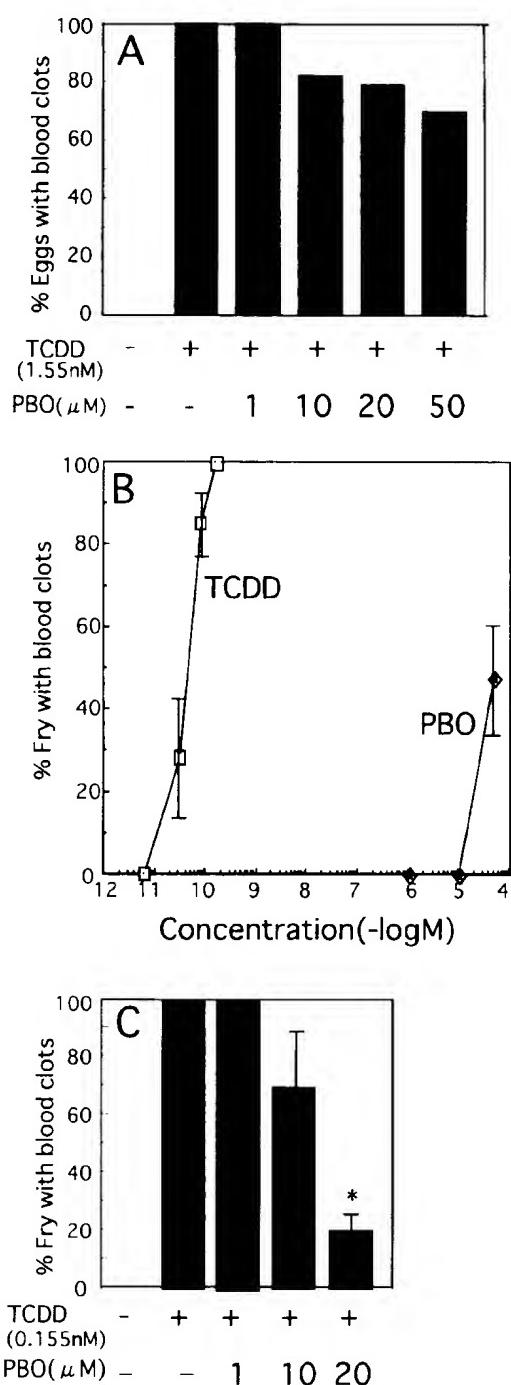


Fig. 4. Suppression by piperonyl butoxide (PBO) of 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD)-induced blood clotting. (A) Eggs were treated with 1.55 nM TCDD and increasing concentrations (μ M) of PBO as indicated until 5 dpf, and examined for blood clotting. (B) Eggs were treated with TCDD and PBO at the indicated concentrations until 5-day post-hatching, and examined for blood clotting in the caudal fin. (C) Eggs were treated with 0.155 nM TCDD and increasing concentrations of PBO as indicated until 5-day post-hatching, and examined for blood clotting in the caudal fin. * $P<0.05$.

TCDD (Fig. 4A). Unexpectedly, PBO reduced the blood clotting rate only slightly; we cannot use higher concentrations of PBO because PBO itself induced blood clotting (described below). We therefore tried to seek for conditions under which lower concentrations of TCDD induce blood clotting effectively. We found that blood clots formed in the caudal fin (Fig. 2Q) after immersing embryos until 5-day post-hatching at subnanomolar concentrations of TCDD (Fig. 4B). Blood clots did not form in the control fin (Fig. 2P). Under the above condition, PBO effectively suppressed the adverse effect of TCDD (Fig. 4C). These results suggest that the TCDD-induced toxicity was caused by elevated expression of a certain Cyt P450.

Previous reports conclude that oxidative stress caused by TCDD-induced expression of Cysts P450 contributes to embryotoxicity and vascular damage associated with apoptosis, because the reducing agent, NAC, partially recovers

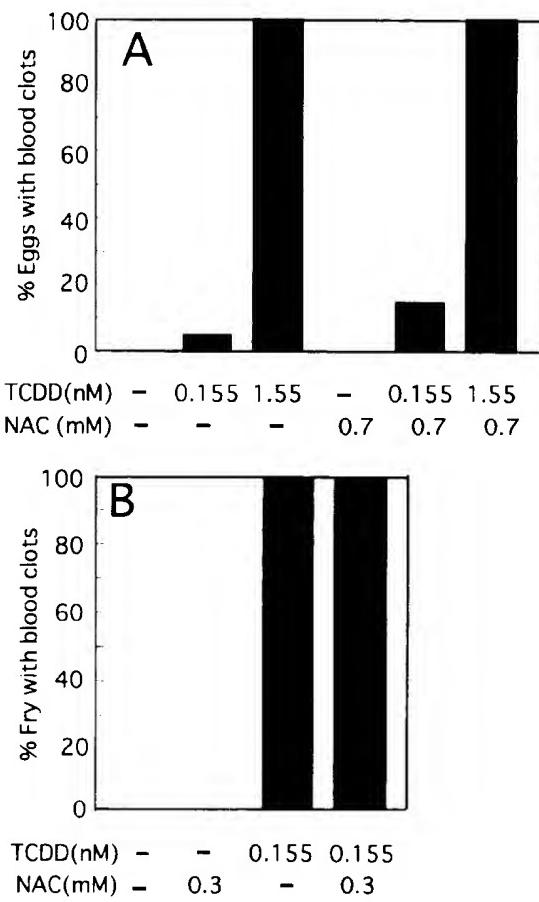


Fig. 5. N-acetyl cysteine (NAC) fails to suppress the 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD)-induced blood clotting. (A) Eggs were treated with TCDD (nM) and NAC (mM) at the indicated concentrations until 5 dpf, and examined for blood clotting. (B) Eggs were treated with 0.155 nM TCDD and 0.3 mM NAC as indicated until 5-day post-hatching, and examined for blood clotting in the caudal fin.

the TCDD-induced embryotoxicity (Cantrell *et al.*, 1996); they observed 41% survival of the embryos that had been treated with 28 nM TCDD for 2 hr and released in 0.1 mM NAC until 3 days posthatch, in contrast to 2% survival of the embryos that had been treated with TCDD and released in water. The ability of NAC to inhibit TCDD-induced toxicity was re-assessed by adding 0.7 mM (Fig. 5A) or 0.3 mM (Fig. 5B) NAC to eggs before and during the treatment with TCDD. NAC could not inhibit the blood clotting induced by 0.155 or 1.55 nM TCDD. NAC itself induced blood clotting at more than 0.9 mM (data not shown). These results suggest that general oxidative stress is not responsible for the TCDD-induced blood clotting.

Vascular damage induced by antagonists (NF and Res) and Cysts P450 inhibitor (PBO)

At the initial experiments determining the concentrations of reagents used, we found that NF, Res, and PBO induced blood clotting at higher concentrations than those used for suppression of TCDD-induced toxicity (Fig. 1). Blood clots formed in caudal and yolk veins (Fig. 2D-F). Yolk veins developed normally at the early time of incubation (up to 4 dpf) (Fig. 2L and N), but their regression was apparent at the time when blood clots formed in yolk veins (at 5 dpf) (Fig. 2M and O). These results suggest that either inactivation of AHR by NF and Res or inhibition of certain Cysts P450 by PBO caused vascular damage and blood clotting.

If the hypothesis were true, antagonist of AHR and Cysts P450 inhibitor would act synergistically to cause toxicity. We examined the synergy between low concentrations of NF (2.5 μ M) and PBO (20 μ M) that alone did not show any effect. Combination of these chemicals clearly increased the

rate of blood clotting (Fig. 6). We therefore conclude that control of AHR activity and levels of Cysts P450 is required for proper development of vasculature in fish.

Malformation or degeneration of bone induced by TCDD, NF, and PBO

During the experiments by incubating eggs with lower concentrations of TCDD (less than 80 pM) until 7 dpf, most eggs developed normally in appearance and blood clots did not form. The eggs were transferred to Yamamoto's solution, then to aquaria after hatching, and reared to adult by normal diet as usual. Unexpectedly we found that these fish were deformed in shape like wavy mutants (Takeuchi, 1960). We examined the bone development by staining with alizarin S. The vertebral column of TCDD-treated fish curved dorso-ventrally and laterally (Fig. 7A and B). Neural and haemal spines were short in length and deformed (Fig. 7B). NF also suppressed the TCDD-induced toxicity on bone formation (Fig. 7C), indicating the involvement of AHR.

We examined the effect of TCDD on the embryonic bone formation by incubating eggs with TCDD until 5 days post-hatching. The staining of the fry with alizarin revealed the absence of calcification in the posterior region of spinal cord and in spines (Fig. 7D and E). We also found that caudal fins were round in shape and constricted (indicated by arrow in Fig. 2S) in the TCDD-treated fry.

In order to examine the possible function of AHR and Cysts P450 in the embryonic bone formation, eggs were treated with NF or PBO until 5 days post-hatching. The treatment with NF (2.5 μ M) did not cause blood clotting in any portion of the fry (data not shown), which was different from the result with TCDD (Figs. 2Q and 4B). However, the treatment also caused degeneration of the posterior end of the spinal cord, but with normal development of spines (Fig. 7D, data not shown). PBO (50 μ M) also caused the same defect in bone formation as that NF did (data not shown).

We further examined whether NF affects homeostasis of adult fish. To do this, adult fish which had been reared by normal diet for 2 months were fed by NF-containing diet (2 mg NF/g diet) for 2 months. During the cultivation, population of fish lacking posterior fins including anal, caudal, and dorsal fins appeared after a month and became increasing near to 100% by two months (Fig. 7F).

Taken together, these results suggest that hyperactivation of AHR by TCDD is toxic to the embryonic development of bone and caudal fin, that AHR is required for proper development of bone and homeostasis of posterior fins, and that a certain Cyt P450 is also required for bone development.

Isolation and characterization of cDNAs encoding AHR homologs of medaka fish, and ubiquitous expression of AHR mRNA

We first obtained four independent cDNA clones (clones 1, 2, 3 and 4) corresponding to PAS domain (Fig. 8A). These clones were found to be identical by sequencing.

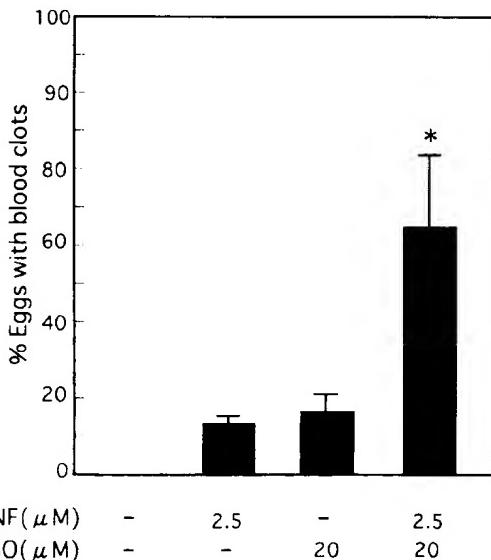


Fig. 6. Synergistic effects of α -naphthoflavone (NF) and piperonyl butoxide (PBO) on blood clotting. Eggs were treated with NF and PBO at the indicated concentrations (μ M) until 6 dpf, and examined for blood clotting. * $P<0.2$.

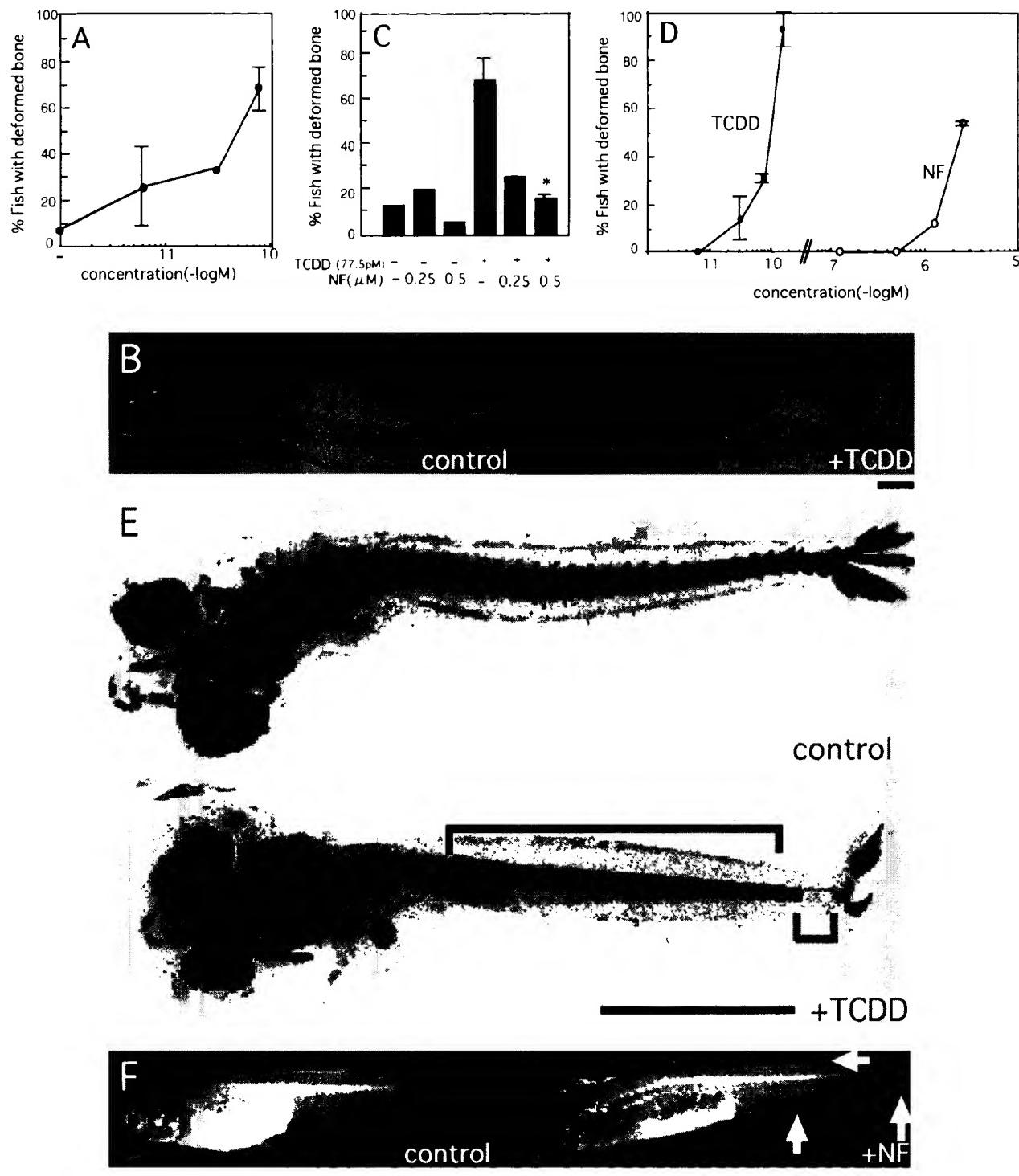


Fig. 7. Effects of 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD) and α -naphthoflavone (NF) on bone formation. (A) Eggs were treated with TCDD at the indicated concentrations until 7 dpf, and reared to adult under TCDD free condition. The adult fish were examined for bone formation after staining with alizarin. (B) Alizarin-stained bone of mock-treated (control) and TCDD (77.5 pM)-treated fish in (A). (C) Eggs were treated with 77.5 pM TCDD and NF at the indicated concentrations (μ M) until 7 dpf, reared to adult under normal condition, and examined for bone formation. * $P<0.05$. (D) Eggs were treated with increasing concentrations of TCDD and NF until 5-day post-hatching, and examined for bone formation. (E) Alizarin stained bone of mock- (control) and TCDD (0.155 nM)-treated fish in (D). Spines and posterior spinal bone are absent in the TCDD-treated fry as noted. (F) Normal adult fish were fed by NF-containing diet (2 mg NF/g diet) for 2 months, and photographed. Arrows indicate the degenerated fins. Bar, 1 mm in (B) and (E), and 5 mm in (F).

Next, 5' and 3' RACEs were performed, yielding four (clones 15, 24, 27 and 30) and six (clones 307–309, 314, 315, and 319) independent clones, respectively (Fig. 8A). Four clones from 5' RACE were identical. Six clones from 3' RACE were subdivided into three identical pairs, which differ from each other only in the 3' proximal sequences denoted by broken and dotted lines in Fig. 8A. Thus, we obtained three different cDNAs, named ahr-1, -2, and -3 (DDBJ accession numbers AB065092, AB065093, and AB065094, respectively). However, ahr-1 and ahr-3 encoded the same protein (AHR1 α), and ahr-2 encoded another homolog (AHR1 β). AHR1 α and

AHR1 β differ from each other in the C-terminal peptides (amino acid 780–879 and 780–784) denoted by shaded and dotted boxes (Fig. 8A).

AHR1 α and AHR1 β are composed of 879 and 784 amino acids with calculated molecular weights of 95.5 and 85.3 kDa, respectively. Both proteins may be classified into a type of AHR1 because they are most homologous to AHR1 of the teleost *Fundulus heteroclitus* (Karchner *et al.*, 1999) (Fig. 8B). The medaka AHR1 α and AHR1 β are also composed of three conserved domains such as basic-helix-loop-helix (bHLH), Per-ARNT-Sim (PAS), and glutamine-rich

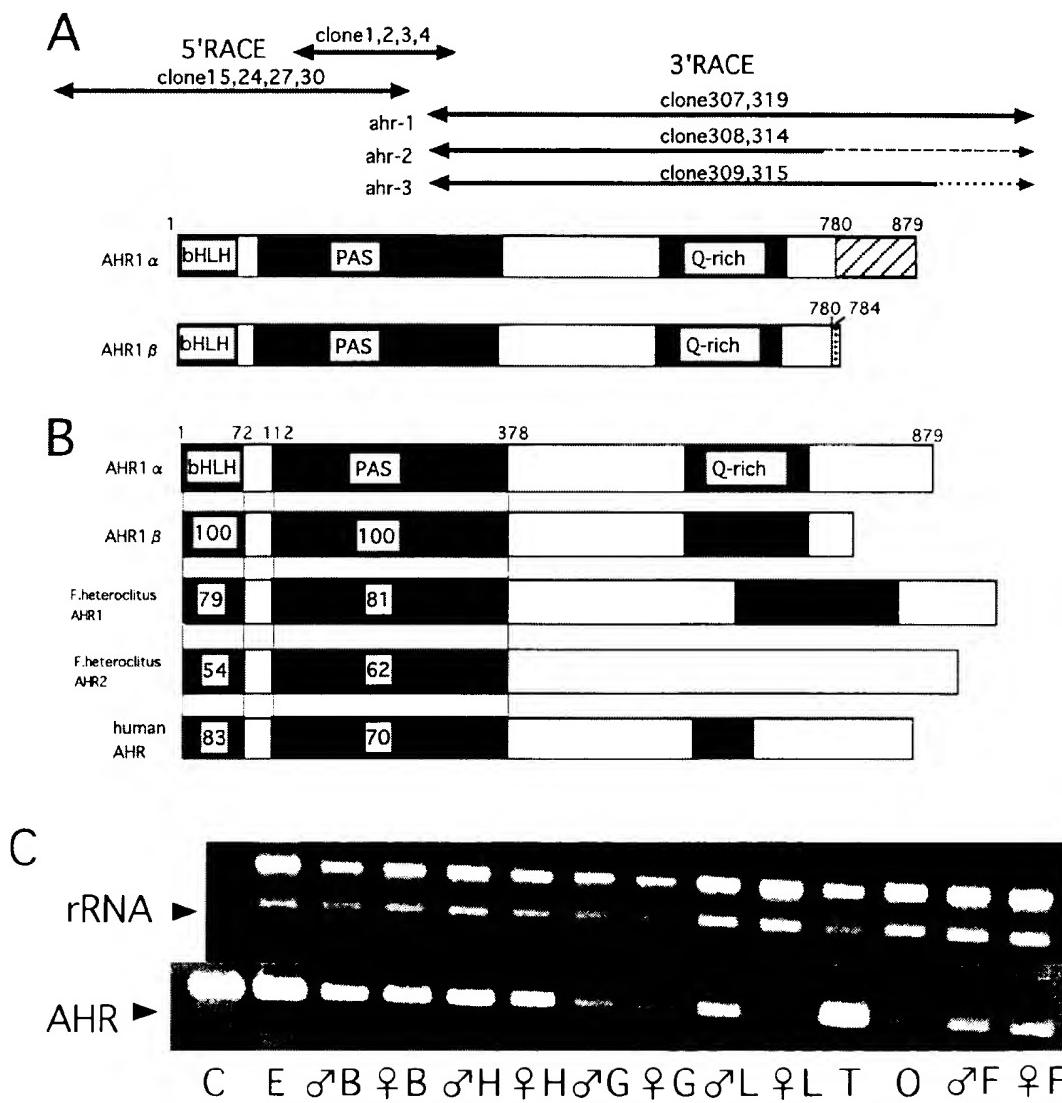


Fig. 8. Schematic drawing of the cDNAs cloned and the deduced proteins, and ubiquitous expression of AHR mRNA. (A) Inserts in the plasmid clones are shown on the deduced proteins (AHR1 α and AHR1 β). Plasmid numbers are marked on the corresponding inserts. The three cDNAs which differ from each other only in the 3' terminal sequences (denoted by broken and dotted lines) are named ahr-1, -2, and -3. AHR1 α and AHR1 β , in which three conserved domains are marked by bHLH, PAS, and Q-rich, differ from each other only in the C-terminal short peptides marked by shaded and dotted boxes. (B) Identity (%) of amino acid sequence among bHLH and PAS domains of AHRs from medaka, *F. heteroclitus* (killifish), and human (Dolwick *et al.*, 1993). (C) RT-PCR analysis of total RNAs from medaka embryos (6 dpf) and adult tissues. Symbols: B, brain; C, the control band amplified from the cDNA; E, embryo; F, caudal fin; G, gill; H, heart; L, liver; O, ovary; and T, testis. Ribosomal RNAs in the RNA samples are also shown.

(Q) domains (Rowlands and Gustafsson, 1997) (Fig. 8B).

Expression of AHR mRNA was analyzed by RT-PCR on total RNAs prepared from medaka embryos and adult tissues such as brain, fin, gill, heart, liver, ovary, and testis. AHR mRNA was detected in all samples tested, and in large amounts in embryos and testis (Fig. 8C).

DISCUSSION

TCDD-induced vascular and bone damages through hyperactivation of AHR

TCDD is the most potent toxicant for vertebrate species. Exposure of vertebrate embryos to TCDD can result in various acute and chronic toxicities such as reproductive failure, teratogenic abnormalities, and immunological dysfunction (Peterson *et al.*, 1993). In fish, vascular damage is the most pronounced adverse effects of TCDD exposure during embryonic development. Vascular hemorrhaging, regression of blood vessels, pericardial sac edema, and reduced circulation are hallmark indicators that vascular function is compromised in the developing embryos (Cantrell *et al.*, 1996; Henry *et al.*, 1997; Hornung *et al.*, 1999; Guiney *et al.*, 2000). The vascular lesions have been demonstrated to be associated with apoptosis and induced expression of Cyt P450 1A in blood vessels of medaka embryos (Cantrell *et al.*, 1998). In the present study, we re-examined the TCDD-induced vascular damage in medaka embryos by observing blood clotting and regression of blood vessels. We found that these vascular damages can be suppressed, but transiently, by antagonist, NF (Fig. 3), giving a convincing evidence that the TCDD-induced vascular damage is mediated through hyperactivation of AHR. The transient suppression may be explained by the fact that TCDD, but not NF, is very stable *in vivo* against catabolic activities of Cysts P450 (Miniero *et al.*, 2001). Although the damage can also be suppressed by Cysts P450 inhibitor, PBO (Fig. 4C), general oxidative stress caused by Cysts P450-mediated oxidative reactions may not be responsible for the

TCDD-induced damage, in inconsistent with the previous conclusion (Cantrell *et al.*, 1996), because reducing agent, NAC, could not recover the damage in vasculature (Fig. 5) or also in bone (data not shown). We assume that a toxic compound that may be accumulated *in vivo* by elevated levels of Cyt P450 is responsible for the TCDD-induced pathology (Fig. 9).

We also found that embryonic treatment with picomolar concentrations of TCDD causes malformation of bone in adult fish (Fig. 7). The treatment did not give apparent complications including blood clotting in the hatching fry, thus, the bone staining is the most sensitive method for detecting TCDD toxicity. The bone deformity could also be recovered by co-treatment with the antagonist (Fig. 7C), implying the role of hyperactivated AHR. TCDD may directly act on bone, because it inhibits osteogenesis in bone-forming cultures of chicken and rat cells (Gierthy *et al.*, 1994; Singh *et al.*, 2000). Treatment of medaka fish with TCDD from the egg stage to post-hatching also caused developmental defects in bone formation at the posterior region of vertebral column and at spines (Figs. 7D, E). However, it may be possible that these defects occurred secondarily to vascular damage, because blood clots formed at the base of the caudal fin under the same condition (Fig. 2Q).

AHR is required for prevention of blood clotting and for proper development of vasculature and bone in medaka fish

AHR is conserved among vertebrates, and ubiquitously expressed in embryos and adult tissues. In the present study, we have cloned three different cDNAs encoding two AHR homologs from medaka fish, *O. latipes* (Fig. 8). The two homologs obtained may belong to a type of AHR1 by amino acid sequence similarity, thus named AHR1 α and AHR1 β . They differ from each other only in C-terminal short peptide, and may be derived from alternative splicing. AHR1 mRNA was also ubiquitously expressed in medaka embryos and adult tissues, suggesting developmental and physiolog-

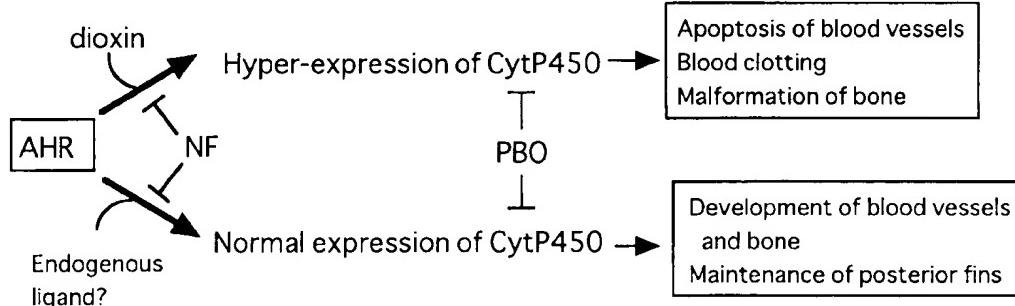


Fig. 9. Model for the role of AHR in the TCDD (dioxin)-induced toxicity, the development of blood vessels and bone, and the maintenance of posterior fins in the medaka fish, *O. latipes*. TCDD-bound AHR induces hyper-expression of a certain Cyt P450, resulting in the toxicities such as apoptosis of blood vessels, blood clotting, and malformation of bone. Either the antagonist (NF) or the Cysts P450 inhibitor (PBO) can suppress the TCDD-induced toxicity. An endogenous ligand is bound to and constitutively activates AHR. The activated AHR is responsible for normal expression of a certain Cyt P450 that is required for the development of blood vessels and bone and homeostasis of posterior fins. *In vivo* inhibition of AHR and Cyt P450 by NF and PBO, respectively, causes developmental abnormalities in vasculature and bone.

ical roles in medaka fish.

To investigate the role of AHR in fish development and physiological homeostasis, medaka embryos (12 hpf) were treated with the antagonists, NF and Res. These compounds did not cause any apparent defects until 4 dpf, but displayed developmental toxicities such as blood clotting and regression of blood vessels at 5 dpf (Figs. 1 and 2). Blood clotting may be caused by regression of blood vessels, because platelet adhesion to subendothelial collagens and activation by components of the extracellular matrix are crucial for blood coagulation (Nieswandt *et al.*, 2001). NF also caused the malformation of bone at 5-day post-hatching (Fig. 7D) and the regression of posterior fins such as anal, caudal, and dorsal fins at the adult period (Fig. 7F). These results suggest the presence of an endogenous ligand for AHR and that constitutive activation of AHR is specifically required for the development of blood vessels and bone and for the maintenance of posterior fins (Fig. 9).

Ligand-bound AHR activates transcription of a battery of genes including various Cyt P450. If levels of a certain Cyt P450 were controlled by AHR bound to an endogenous ligand and required for proper development of blood vessels and bone, the well-known inhibitor (PBO) of the enzymatic activity of Cyt P450 would induce the same developmental defect as did the antagonist. Treatment of embryos with PBO specifically induced blood clotting, regression of blood vessels (Figs. 1 and 2), and degeneration of the posterior end of spinal cord (data not shown) at the same developmental stage as did the antagonist, suggesting the importance of Cyt P450, the identity of which is, however, unknown (Fig. 9). The synergistic effects exerted by NF and PBO (Fig. 6) also support the hypothesis. We assume that a certain Cyt P450 is responsible for degradation (or catabolism) of a toxic compound that caused the developmental abnormalities.

ACKNOWLEDGEMENTS

We thank S. Omura and R. Horie for assistance and discussion. This study was supported in part by a grant from Iyaku-shigen Research Foundation.

REFERENCES

- Abbott BD, Schmid JE, Pitt JA, Buckalew AR, Wood CR, Held GA, Dilberto JJ (1999) Adverse reproductive outcomes in the transgenic Ah receptor-deficient mouse. *Toxicol Appl Pharmacol* 155: 62–70
- Adams NH, Levi PE, Hodgson E (1993) Regulation of cytochrome P-450 isozymes by methylenedioxynaphthalene compounds. *Chem Biol Interact* 86: 255–274
- Cantrell SM, Joy-Schlezinger J, Stegeman JJ, Tillitt DE, Hannink M (1998) Correlation of 2,3,7,8-tetrachlorodibenzo-p-dioxin-induced apoptotic cell death in the embryonic vasculature with embryotoxicity. *Toxicol Appl Pharmacol* 148: 24–34
- Cantrell SM, Lutz LH, Tillitt DE, Hannink M (1996) Embryotoxicity of 2,3,7,8-tetrachlorodibenzo-p-dioxin (TCDD): The embryonic vasculature is a physiological target for TCDD-induced DNA damage and apoptotic cell death in medaka (*Oryzias latipes*). *Toxicol Appl Pharmacol* 141: 23–34
- Casper RF, Quesne M, Rogers IM, Shirota T, Jolivet A, Milgrom E, Savouret JF (1999) Resveratrol has antagonist activity on the aryl hydrocarbon receptor: Implications for prevention of dioxin toxicity. *Mol Pharmacol* 56: 784–790
- Ciolino HP, Daschner PJ, Yeh GC (1998) Resveratrol inhibits transcription of CYP1A1 *in vitro* by preventing activation of the aryl hydrocarbon receptor. *Cancer Res* 58: 5707–5712
- Dahl AR, Hodgson E (1979) The interaction of aliphatic analogs of methylenedioxynaphthalene compounds with cytochromes P-450 and P-420. *Chem Biol Interact* 27: 163–175
- Dolwick KM, Schmidt JV, Carver LA, Swanson HI, Bradfield CA (1993) Cloning and expression of a human Ah receptor cDNA. *Mol Pharmacol* 44: 911–917
- Fernandez-Salguero PM, Hilbert DM, Rudikoff S, Ward JM, Gonzalez FJ (1996) Aryl-hydrocarbon receptor-deficient mice are resistant to 2,3,7,8-tetrachlorodibenzo-p-dioxin-induced toxicity. *Toxicol Appl Pharmacol* 140: 173–179
- Fernandez-Salguero P, Pineau T, Hilbert DM, McPhail T, Lee SS, Kimura S, Nebert DW, Rudikoff S, Ward JM, Gonzalez FJ (1995) Immune system impairment and hepatic fibrosis in mice lacking the dioxin-binding Ah receptor. *Science* 268: 722–726
- Gasiewicz TA, Rucci G (1991) Alpha-naphthoflavone acts as an antagonist of 2,3,7,8-tetrachlorodibenzo-p-dioxin by forming an inactive complex with the Ah receptor. *Mol Pharmacol* 40: 607–612
- Gierthy JF, Silkworth JB, Tassinari M, Stein GS, Lian JB (1994) 2,3,7,8-Tetrachlorodibenzo-p-dioxin inhibits differentiation of normal diploid rat osteoblasts *in vitro*. *J Cell Biochem* 54: 231–238
- Gonzalez FJ, Fernandez-Salguero P (1998) The aryl hydrocarbon receptor. *Drug Metab Dispos* 26: 1194–1198
- Guiney PD, Smolowitz RM, Peterson RE, Stegeman JJ (1997) Correlation of 2,3,7,8-tetrachlorodibenzo-p-dioxin induction of cytochrome P4501A in vascular endothelium with toxicity in early life stages of lake trout. *Toxicol Appl Pharmacol* 143: 256–273
- Guiney PD, Walker MK, Spitsbergen JM, Peterson RE (2000) Hemodynamic dysfunction and cytochrome P4501A mRNA expression induced by 2,3,7,8-tetrachlorodibenzo-p-dioxin during embryonic stages of lake trout development. *Toxicol Appl Pharmacol* 168: 1–14
- Hahn ME, Karchner SI (1995) Evolutionary conservation of the vertebrate Ah (dioxin) receptor: amplification and sequencing of the PAS domain of a teleost Ah receptor cDNA. *Biochem J* 310: 383–387
- Hankinson O (1995) The aryl hydrocarbon receptor complex. *Annu Rev Pharmacol Toxicol* 35: 307–340
- Henry TR, Spitsbergen JM, Hornung MW, Abnet CC, Peterson RE (1997) Early life stage toxicity of 2,3,7,8-tetrachlorodibenzo-p-dioxin in zebrafish (*Danio rerio*). *Toxicol Appl Pharmacol* 142: 56–68
- Hornung MW, Spitsbergen JM, Peterson RE (1999) 2,3,7,8-tetrachlorodibenzo-p-dioxin alters cardiovascular and craniofacial development and function in sac fry of rainbow trout (*Oncorhynchus mykiss*). *Toxicol Sci* 47: 40–51
- Karchner SI, Powell WH, Hahn ME (1999) Identification and functional characterization of two highly divergent aryl hydrocarbon receptors (AHR1 and AHR2) in the teleost *Fundulus heteroclitus*. *J Biol Chem* 274: 33814–33824
- Kawahara T, Okada H, Yamashita I (2000) Cloning and expression of genomic and complementary DNAs encoding an estrogen receptor in the medaka fish, *Oryzias latipes*. *Zool Sci* 17: 643–649
- Kawahara T, Yamashita I (2000) Estrogen-independent ovary formation in the medaka fish, *Oryzias latipes*. *Zool Sci* 17: 65–68
- Merchant M, Krishnan V, Safe S (1993) Mechanism of action of alpha-naphthoflavone as an Ah receptor antagonist in MCF-7

- human breast cancer cells. *Toxicol Appl Pharmacol* 120: 179–185
- Mimura J, Yamashita K, Nakamura K, Morita M, Takagi TN, Nakao K, Ema M, Sogawa K, Yasuda M, Katsuki M, Fujii-Kuriyama Y (1997) Loss of teratogenic response to 2,3,7,8-tetrachlorodibenzo-*p*-dioxin (TCDD) in mice lacking the Ah (dioxin) receptor. *Genes Cells* 2: 645–654
- Miniero R, De Felip E, Ferri F, di Domenico A (2001) An overview of TCDD half-life in mammals and its correlation to body weight. *Chemosphere* 43: 839–844
- Nieswandt B, Brakebusch C, Bergmeier W, Schulte V, Bouvard D, Mokhtari-Nejad R, Lindhout T, Heemskerk JWM, Zirngibl H, Faessler R (2001) Glycoprotein VI but not $\alpha 2\beta 1$ integrin is essential for platelet interaction with collagen. *EMBO J* 20: 2120–2130
- Peterson RE, Theobald HM, Kimmel GL (1993) Developmental and reproductive toxicity of dioxins and related compounds: cross-species comparisons. *Crit Rev Toxicol* 23: 283–335
- Robles R, Morita Y, Mann KK, Perez GI, Yang S, Matikainen T, Sherr DH, Tilly JL (2000) The aryl hydrocarbon receptor, a basic helix-loop-helix transcription factor of the PAS gene family, is required for normal ovarian germ cell dynamics in the mouse. *Endocrinol* 141: 450–453
- Rowlands JC, Gustafsson JA (1997) Aryl hydrocarbon receptor-mediated signal transduction. *Crit Rev Toxicol* 27: 109–134
- Singh SUN, Casper RF, Fritz PC, Sukhu B, Ganss B, Girard Jr B, Savouret JF, Tenenbaum HC (2000) Inhibition of dioxin effects on bone formation *in vitro* by a newly described aryl hydrocarbon receptor antagonist, resveratrol. *J Endocrinol* 167: 183–195
- Takeuchi K (1960) A study of the mutant (*wavy*) in the medaka, *Oryzias latipes*. *Annotationes Zoologicae Japonenses* 33: 124–131
- Testa B, Jenner P (1981) Inhibitors of cytochromes p450 and their mechanisms of action. *Drug Metab Rev* 12: 1–117
- Yamamoto T (1969) Sex differentiation. In "Fish Physiology 3" Ed by WS Hoar, DJ Randall Academic Press, New York, pp 117–175

(Received September 12, 2001 / Accepted December 3, 2001)

[TOP](#), [HOME](#)

Tokio Yamamoto In: "Medaka, Biology and Strains" (T. Yamamoto, ed.), Yugakusya Publ. (1975), pp. 17-29.

Systematics and Zoogeography

The killifishes, or Cyprinodontiforms are small fresh and brackish water fishes of worldwide distribution in tropical and temperate latitudes.

Previous classification of the order Cyprinodontes

The classification of the order Cyprinodontes Agassiz (equivalent to Microcyprini Regan) has been worked out by Gill (1865, 1874), Regan (1909, 1911), Hubbs (1924, 1926) and Myers (1931, 1938). The classification followed here is mostly according to Hubbs and Myers and is cited from Kulkalni (1940) who erected a new family Horaichthyidae represented by a remarkable Indian henpecked killifish, *Horaichthys setnai*. However, substituting for the terms Amblyopsoidea and Poecilioidea, the suborders Amblyopsoidei and Cyprinodontoidei are used here, respectively. The subfamily Tomeurinae is removed from the family Poeciliidae to erect a new family Tomeuridae as suggested by Hubbs in his letter to S. L. Hora (India) in 1938. Representative genera are given in parentheses following family names.

Order Cyprinodontes (Microcyprini)

Suborder Amblyopsoidei

Family Amblyopsidae (*Chologaster, Amblyopsis*)

Suborder Cyprinodontoidei

Family Cyprinodontidae (*Cyprinodon, Fundulus,*

Aplocheilus, Panchax, Oryzias)

Family Goodeidae (*Goodea*)

Family Poeciliidae (*Poecilia, Gambusia, Xiphophorus*)

Family Jenynsiidae (*Jenynsia*)

Family Anablepidae (*Anableps*)

Family Tomeuridae (*Tomeurus*)

Family Adrianichthyidae (*Adrianichthys, Xenopoecilus*)

Family Phallostethidae (*Phallostethus, Gulaphallus*)

Family Horaichthyidae (*Horaichthys*)

The classification listed here has been generally held by ichthyologists until 1962.

As to the status of *Oryzias*, Myers (1931) considered it to represent a monogeneric tribe of the subfamily Fundulinae. Later (1956), he revised his earlier classification, and considered *Oryzias* to represent a monogeneric subfamily of the Cyprinodontidae, the Oryziatinae.

Classification of new order Atherinoformes

Rosen (1962) presented evidence which indicates a relationship of the Amblyopsidae (North American cave fishes) with the percopsiform genera and, more distantly with the gadiforms. He isolated the cave fishes as a new order, the Amblyopsiformes, and recommended its alignment near the Percopsiformes and Gadiformes in a phyletic sequence.

In 1964, Rosen has made drastic taxonomic re-arrangements of the halfbeaks, killifishes, silversides, and their relatives. The outset of his re-arrangements was osteological analyses of the adrianichthyid fishes of Celebes, which were found to have a mixture of beloniform, cyprinodontiform, and mugiliform features. Then, his investigation was broadened to include representatives of all these groups as well as a species of phallostethid.

In consequence of reasonable osteological diagnoses, he erected a new order Atherinoformes which includes the exocoetoids, scomberesocoids. On the basis of osteological evidence, he separated the medaka (*Oryzias*) from cyprinodontoids, placed it in adrianichthyoids and erected a new family Oryziatidae.

To visualize Rosen's account on osteological difference between cyprinodontoids and adrianichthyoids, the presentation of the schema of the skull of the generalized teleosts as shown in Fig. 2-1 may be apropos.

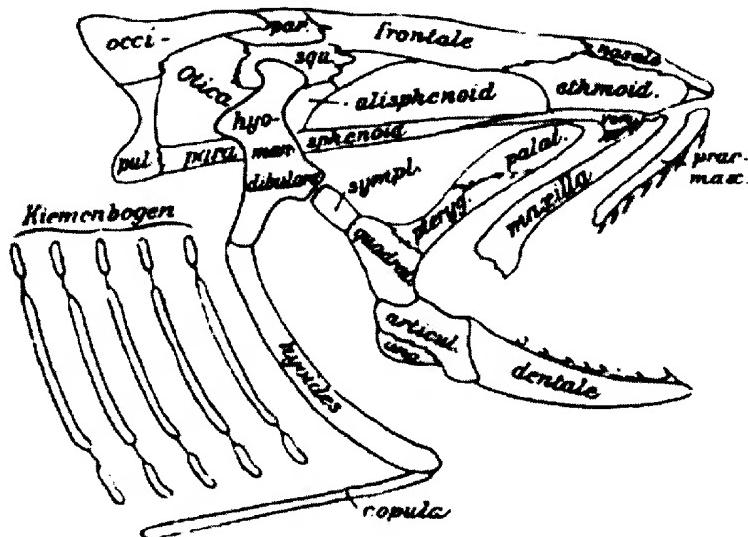


Fig. 2-1. A diagram of teleostean skull. Opercula and Infraorbitalia are removed. ang. = angular, articul. = articular, occi. = occipital, palat. = palatine, p = quadrate, squ. = squamosa, sympl. = symplectic, vom. = vomer.

After R. Goldschmidt' E. Selenkas Zoologishes Taschenbuch fur Studierende. 1912 Leipzig, George Thieme.

In cyprinodontoid killifishes, bones of the jaws and the palatoquadrate arch are in such a construction that the premaxilla is protractile. In adrianichthyoid killifishes, on the other hand, the premaxillae are not protractile. The Adrianichthyidae are fishes of small size confined to the fresh-water lakes of Celebes. Two species, *Adrianichthys kruyti* and *Xenopoecilus sarasinorum*, are known. *Xenopoecilus* is characterized by having a large horse-shoe shaped mouth, an enormous ethmoideum and a single, median supraoccipital process formed by fusion of embryologically paired elements; "a cup-like excavation on the distal tip of the autopatinate that is capped by a large ball of cartilage and a discoidal sesamoid bone; a dorsal enlargement of the palatoptygoid arch with a prefrontal (Fig. 2-2); a maxilla that is carried on the upper edge rather than on the outer face of the posterior end of the premaxilla; a premaxilla that lacks a hooked or pointed posteroventral process; a tremendously reduced articular bone without a coronoid process that is almost wholly contained within the posterior part of the dentary; the articulation of the first pleural rib on the third rather than on the second vertebra; pelvic girdles that are not in contact medially and that have a long lateral spur extending upward between ribs; a dorsoventrally asymmetrical caudal skeleton with one or two very slender, rod-like epurals, and a caudal fin that is divided into indistinct upper and lower lobes by having a large gap between rays that articulate with the upper and lower hypural plates on the terminal half-centrum. (Rosen, 1964)

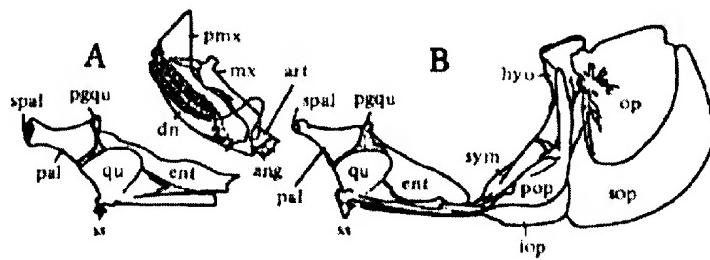


Fig. 2-2. Jaws and jaw suspension in adrianichthyoid killifishes. A. Jaws and palatoptyrgoid arch in *Oryzias latipes* (Temminck and Schlegel). b. Jaw suspension and opercular apparatus in *Xenopoecilus sarasinorum* (Popa). Note sesamoid bone below quadrate and bony cap over tip of palatine in A. and B. Note in A that lower arm of premaxilla lies over maxilla, large coronoid process on dentary, and absence of similar coronoid elevation on articular. Ang = angular, art = articular, dn = dentary, ent = entpterygoid (mesopterygoid), hyo = hyomandibular, iop = interoperculum, mx = maxilla, op = operculum, pal = palatine (autopalatine with or without dermopalatine), pgqu = pterygoquadrate cartilage, pmx = sesamoid bone capping autoplatine, ss = sesamoid bone, sym =symplectic. Rosen, 1964.

Rosen pointed out that except for the enlarged jaws and the presence of a median supraoccipital process, all the above features described within quotation marks can be identified in *Oryzias* (Fig. 2-2) but in no other killifishes so far as known.

It is therefore apparent that adrianichthyids and the medaka are intimately related and that they constitute a distinct subgroup of the killifishes, the adrianichthyoids, containing the families Adrianichthyidae (*Adrianichthys* and *Xenopoecilus*), Oryziatidae (*Oryzias*), and Horaichthyidae (*Horaichthys*), in contrast to the remainder of the families which are grouped together as cyprinodontoids (Cyprinodontoidea). Basing on Rosen's (1964) findings, Turner (1965) conveniently enumerated difference between cyprinodontoids and *Oryzias* as follows:

Cyprinodontidae	<i>Oryzias</i>
1. First pleural rib on second vertebra.	First pleural rib on third vertebra.
2. Pelvic girdle bones joined mid-ventrally; no upright lateral spur.	Pelvic girdle bones not joined mid-ventrally; an upright lateral spur present.
3. Lower end of premaxilla bone expanded or hooked and sandwiched between the lower end of maxilla bone and dentary bone (lower jaw).	Lower end of premaxilla bone not expanded, and dorsal to the maxilla bone rather than between it and the dentary bone.
4. Hypural plates often fused.	Hypural plates never fused.
5. Hypochordal musculature entirely absent.	Hypochordal musculature present.
6. Caudal fin never incipiently lobed.	Caudal fin incipiently lobed.

The family Horaichthyidae erected by Kulkarni (1940) comprises a single species, *Horaichthys setnai*. It is a small translucent oviparous fish inhabiting brackish waters and estuaries in the province of Bombay, India. Osteological study (Kulkalni 1948) showed that its head skeleton is closely allied to that of *Oryzias* but greatly different from that of *Aplocheilus*. *Horaichthys*, however, is different from known species of *Oryzias* in having a larger number of the anal fin- rays (about 28 to 32).

In the male, six anterior rays of the anal fin are separated from the rest of the fin and modified into an elaborate male organ (gonopodium). Of six rays the third, fourth and fifth ones are profoundly modified forming the 3- 4-5 complex. (Fig. 2-3). In the female right pelvic fin is usually absent. The genital opening of the female is situated on the left ventral side and is surrounded by genital pads. *Horaichthys* is supposed to have evolved from *Oryzias*, but as the development of the gonopodium in association with the henpecked sexual behavior is so remarkable that Kulkarni (1940) has proposed to erect a new family rank for this fish.

The male appears to be always afraid of the female which on occasions chases him away. At the time of mating, "the male swims below and behind her at a distance of about 2 to 3 cm. He then darts towards her on the left with almost lightning speed. As he approaches his mate he lashes out the gonopodium sideways almost at right angles to his body and strikes its terminal end against her genital opening. The spermatophores are transferred to the female in this momentary contact, and become attached by their distal hooks."

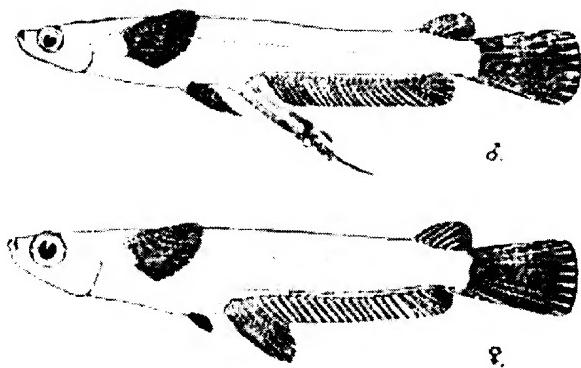


Fig. 2-3. Lateral view of a male and a female specimen of *Horaichthys setnai*.
x 4 Kulkalni, 1940.

A special feature of *Horaichthys* is that the testis produces special sperm capsules of spermatophores (2-300 in number) instead of ordinary semi-fluid milt with suspended sperms.

A spermatophore is a tiny hyaline body (0.6 mm long and 0.1 mm thick), the broad part of which contains mass of sperms. At the tapering end, there is a pointed cap with stiff hooks and barb-like structures which point backwards. It is with the aid of these hooks and barbs that the spermatophore get attached near the genital opening of the female.

There is no permanent opening on the spermatophore for the liberation of sperms. Before liberation of sperms, a small bulging appears at the neck of the tapering spermatophore and begins to enlarge. When the protuberance becomes sufficiently large, an opening is formed at its tip by rupture of membrane and sperms are liberated. They swim into the genital pore of the female.

The following is the classification of the new order Atheriniformes by Rosen (1964), representative species being given in parentheses.

Suborder Exocoetoidei

Superfamily Exocoetoidea

Family Hemiramphidae (*Hemiramphus*)

Family Exocoetidae (*Exocoetus*)

Superfamily Scomberesocoidea

Family Belonidae (*Ablennus*)

Family Scomberesocidae (*Cololabis*)

Suborder Cyprinodontoidei

Superfamily Adrianichthyoidea

Family Oryziatidae (*Oryzias*)

Family Adrianichthyidae (*Adrianichthys, Xenopoecilus*)

Family Horaichthyidae (*Horaichthys*)

Superfamily Cyprinodontoidea

Family Cyprinodontidae (*Fundulus*, *Aplocheilus*)
Family Goodeidae (*Goodea*)
Family Jenynsiidae (*Jenynsia*)
Family Anablepidae (*Anableps*)
Family Poeciliidae (*Poecilia*, *Xiphophorus*)
Suborder Atherinoidei
Superfamily Atherinoidea
Family Melanotaeniidae
Family Atherinidae (*Atherina*)
Family Isonidae, new family (*Iso*)
Superfamily Phallostethoidea
Family Neostethidae (*Neostethus*)
Family Phallostethidae (*Phallostethus*)

The family Oryziatidae

Rosen (1964) erected a new monogeneric family and described the following diagnoses of the family Oryziatidae. Type genus: *Oryzias* Jordan and Snyder, 1906. Diagnoses: The Oryziatidae differ from their closest relatives, the adrianichthyids, in lacking the tremendously enlarged jaws and ethmoideum, in having paired supraoccipital processes (rather than a single median process), and in having the inferior pharyngeal bone distinctly separated (rather than united), and from all cyprinodontoids as follows: autopalatine usually capped by sesamoid bone; pterygoquadrate cartilage forming dorsal process; lower end of premaxilla not hooked or trapezoidal, situated below maxilla rather than between maxilla and dentary bone; first pleural rib on third vertebra; supracleithrum wanting; pelvic bones with upright lateral spurs and not joined midventrally; hypochordal musculature present on caudal fin.

Composition: Rosen listed following seven species of a single genus, *Oryzias*: *O. latipes* (Temminck and Schlegel), *O. melastigma* (McClelland), *O. celebensis* (Weber), *O. timorensis* (Weber and de Beaufort), *O. javanicus* (Bleeker), *O. curvinotus* (Nichols and Pope), and *O. minutillus* Smith. To these, *O. luzonensis* (Herré and Ablan) may be added. Besides these, Turner (1965) mentioned *O. matenensis* (Aurich), and *O. marmoratus* (Aurich) from the Celebes.

Probably not all these nominal species are valid, since some nominal species are different only in the anal fin-ray frequency.

The genus *Oryzias*

The following is the diagnoses of the genus *Oryzias* described by Jordan and Snyder (1906), basing on *O. latipes* which has previously been known as *Aplocheilus latipes*.

Body elliptical in form, compressed, covered with large scales; mouth small, with two rows of small, simple, pointed teeth; *no teeth on vomer**1; gill-opening not restricted above; intestinal canal short, about as large as body; peritoneum black. Dorsal fin short, inserted above middle of anal; anal very long seventeen to twenty rays; caudal fin truncate. *Sexes similar**2 *except color*; anal fin not modified in the male. *1 Kulkarni(1948) first showed that *os vomer* is absent in *Oryzias melastigma*. *2 Sexual

dimorphism is prominent. See Chap. 8.

The species *Oryzias latipes*

The following description by Oshima (1919) based on a specimen of *Oryzias latipes* collected from Shori, Formosa is cited here as the diagnoses of the species since it is very precise and correct excepting two words starred and daggered.

Head 4 in length (body length divided by head length is 4); depth 4.5; depth of caudal peduncle 9.5; eye 2.5 in head (head length divided by eye diameter is 2.5); interorbital space 2; snout 4; D.6; A.18; P.9; V.5; thirty one scales in a lateral series; five branchiostegals.

Posterior half of the body compressed, becoming broader anteriorly, highest in front of the anal; head flattened; interorbital space broad; snout shorter than the diameter of eye, broadly rounded anteriorly; mouth anterior, transverse; lower jaw slightly projecting, each jaw with two rows of minute pointed teeth, those on the posterior row smaller; vomer*1 smooth; thirteen short, pointed gill-rakers on the first arch; eyes very large, anterior and superior.

Dorsal fin short, on the posterior half of body, its origin above the posterior two thirds of anal, its height equal to the distance between tip of snout and posterior margin of orbit; pectoral inserted on the median line of body; the ventral small, reaching vent; base of the anal very long, its posterior end opposite to that of the dorsal, anterior ray longest; tip of the caudal fin *rounded*.*2

Top and sides of head, throat, and chin naked; body covered with cycloid scales, lateral line absent.

Color in formalin pale gray above, lower parts silvery; a black longitudinal streak from the nape to the origin the dorsal; sides of body with a faint dusky stripe along the middle line, top of head dark; the edges of scales dusky; fin-rays of the ventral and anal dotted with minute black spots; all the fins whitish; peritoneum black. Length of body 28 mm.

Habitat: The present species is very common in rice-fields and pools on the island.

*1 Vomer is absent in *Oryzias* in reality.

*2 The caudal fin is almost truncate, strictly, however, it is incipiently lobed.

Change of nomenclature of the medaka

The medaka was first described as *Poecilia latipes* by Temminck and Schlegel in 1846 (Siebold's Fauna Japonica, Poiss., P.224, Pl.102, Fig. 5). Günther changed it as *Haplochilus latipes*. Jordan and Snyder (1901) described it as *Apolochilus latipes* but later they separated it from *Apolochilus* and erected a new genus *Oryzias*. They regarded *Oryzias* as having no teeth on vomer*1 while *Apolochilus* possesses teeth on it.

Myers (1931) placed the medaka in the tribe Aplocheilini of the subfamily Fundulinae in the family Cyprinodontidae. He stated that the chief character of fishes of the tribe is the non-protractile premaxillae. The pectoral fin are set high and pseudobranchiae and vomerine teeth are never present. The species range from Japan and Central China south to Celebes and Timor and west to Southern India. A single genus, *Aplocheilus*, of which *Oryzias* is a synonym*2. Smith, (1945) pointed out that the genus known as Panchax is a synonym of *Aplocheilus* McClelland and *Aplocheilus* Weber and de Beaufort is a synonym of *Oryzias* Jordan and Snyder. He described *Aplocheilus panchax* (Hamilton) and *Oryzias minutillus* n. sp. from Thailand.

According to him, the two genera may be distinguished by the following characters:

- a. Upper jaw protractile; mouth moderate size with its corners abruptly bent downward; vomer toothed; pseudobranchiae present; branchial membranes free from each other and from isthmus; pectoral fins with their upper base at or below longitudinal axis of body Aploc
 - b. Upper jaw not protractile; mouth small with its corners obtusely bent downward; vomer toothless; no pseudobranchiae; branchiae membranes united across isthmus; pectoral fins with upper base well above longitudinal axis of body Oryzia

The correct scientific name of the medaka is *Oryzias latipes* (Temminck and Schlegel).

From Jordan and Snyder (1906) onwards, all taxonomists stated that *Oryzias* has toothless vomer while *Aplocheilus* has toothed vomer. Kulkarni (1948) has made a precise osteological study of Indian killifishes and found that vomer is absent in both *Oryzias melastigma* and *Horaichthys setnai* while *Aplocheilus lineata* possesses toothed vomer.

*1 Vomer is absent in *Oryzias* in reality.

*2 Now, the two genera belong to the different super-families.

Geographical distribution of species belonging to the Genus *Oryzias*

All the species of the genus *Oryzias* are distributed in India, South Asia, the Indo-Australian archipelago and the Far East. Their habitats are widely ranged from tropical, subtropical, and temperate regions as shown in Figure 2-4 and in the following lines.

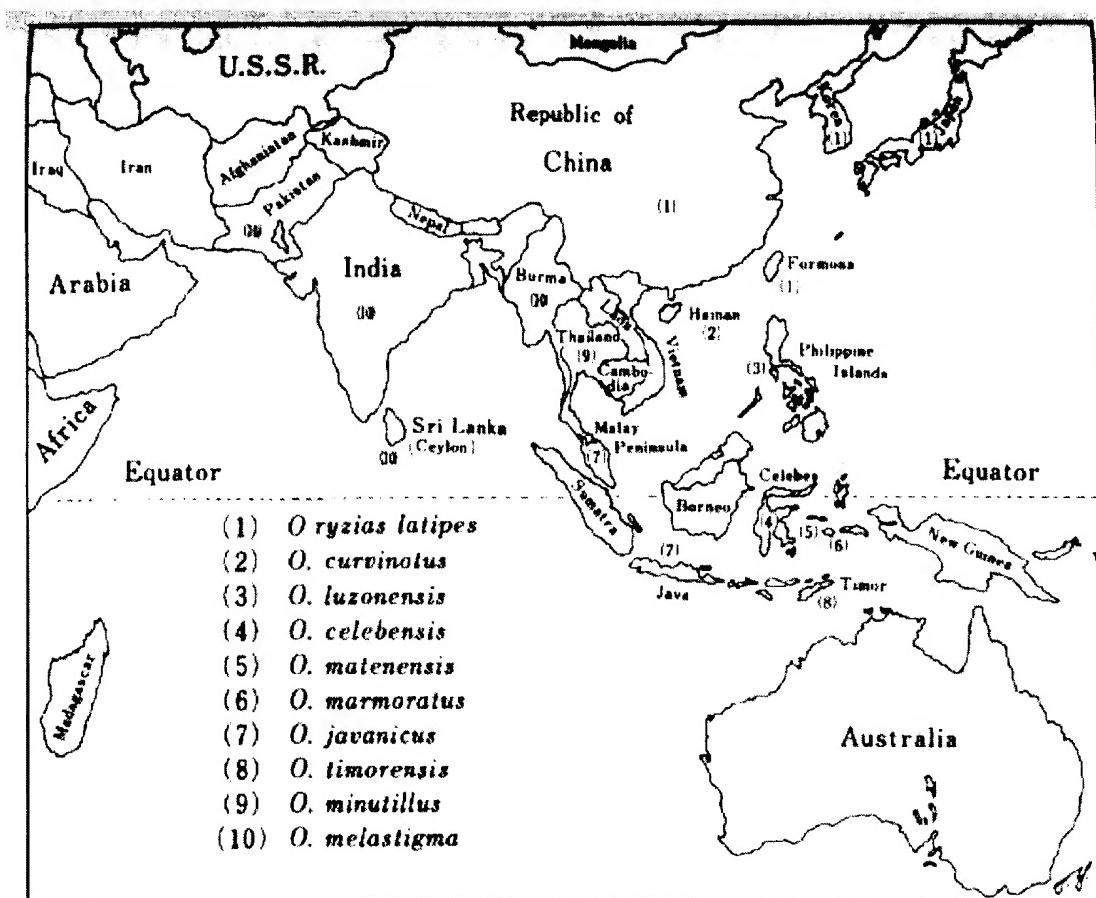


Fig. 2-4. A

zoogeographical map showing distribution of species of the Genus *Oryzias*. Original.

- (1) *O. latipes* (Temminck and Shlegel): Japan, Korea, Formosa, and China
- (2) *O. curvinotus* (Nicols and Pope): The island of Hainan.
- (3) *O. luzonensis* (Herre and Abilan): Luzon in the Philippines.
- (4) *O. celebensis* (Weber): The Celebes.
- (5) *O. matenensis* (Aurich): The Celebes.
- (6) *O. marmoratus* (Aurich): The Celebes.
- (7) *O. javanicus* (Bleeker): The Indo-Malaysian archipelago and Malaya.
- (8) *O. timorensis* (Weber and de Beaufort): The island of Timor.
- (9) *O. minutillus* Smith: Thailand.
- (10) *O. melastigma* (McClelland): India, Western Pakistan, and Sri Lanka (Ceylon).

In the main, all the *Oryzias* species are fresh-water fishes. *O. latipes* and *O. melastigma* inhabit both fresh and brackish water. *O. latipes* is so tolerate salinity that it thrives in tide pools in Korea and Kyushu in Japan.

References

- Gill, T.N., 1965 Synopsis of the fishes in the Gulf of St. Lawrence and Bay of Fundy. Canadian Nat., Ser. 2, 2: (No.4) 244-266.
 Gill, T.N., 1874 Arrangement of the families of fishes, or Classes Pisces, Marssipobranchii and Leptocardii. Smithsonian Misc. Coll., for 1872, 11: (No.247) 1-49.

- Herre, A.W., and G.L. Abian, 1934 *Aplocheilus luzonensis*, a new Philippine Cyprinodont. Philippine Jour. Sci., 54: (No.2) 275-277.
- Hubbs, C.L., 1924 Studies on the fishes of the order Cyprinodontes I.-IV. Misc. Publ. Mus. Zool., Univ. Michigan, No. 13: 1-31.
- Hubbs, C.L., 1926 Studies on the fishes of the order Cyprinodontes VI. Misc. Publ. Mus. Zool., Univ. Michigan, No. 16: 1-87.
- Jordan, D.S., and J.O. Snyder, 1906 A review of the Poeciliidae or Killifishes of Japan. Proc. U.S. Nat. Mus., 31: 287-290.
- Kulkarni, C.V., 1940 On the systematic position, structural modifications, bionomics and development of a remarkable new family of cyprinodont fishes from the province of Bombay. Rec. Indian Mus., 42: 379-423.
- Kulkarni, C.V., 1948 The osteology of Indian cyprinodonts. Part. I. comparative study of the head skeleton of *Aplocheilus*, *Oryzias* and *Horaichthys*. Proc. Natl. Inst. Sci. India, 14: (No.2) 65-119.
- McClelland, J., 1839 Asiatic researches, 19: 301.
- Myers, G.S., 1931 The primary groups of oviparous cyprinodont fishes. Stanford Univ. Publ. Biol. Sci. VI. No. 3: 7-14.
- Myers, G.S., 1938 Studies on the genera of cyprinodont fishes. Copeia (1938): 136-143.
- Nichols, J.T., and C.H. Pope, 1927 The fishes of Hainan. Bull. Amer. Mus. Hist., 54: 321.
- Oshima, M., 1919 Contributions to the study of the fresh water fishes of the island of Formosa. Annals Carnegie Mus., 12: 169-328.
- Regan, C.T., 1909 The classification of teleostean fishes. Ann. Mag. Hist., Ser. 8, 3: 75-86.
- Regan, C.T., 1911 The osteology and classification of the teleostean fishes of the order Microcyprini. Ann. Mag. Nat. Hist., Ser. 8, 7: 320-327.
- Rosen, D.E., 1962 Comments on the relationships of the North American cave fishes of the family Amblyopsidae. Amer. Mus. Novitates, No. 2109: 1-35.
- Rosen, D.E., 1964 The relationships and taxonomic position of the halfbeaks, killifishes, silversides, and their relatives. Bull. Amer. Mus. Nat. Hist., 127: (Art.5) 217-268.
- Smith, H.M., 1938 Status of the oriental fish genera *Aplocheilus* and *Panchax*. Proc. Biol. Soc. Washington, 51: 165-166.
- Smith, H.M., 1945 The fresh-water fishes of Siam, or Thailand. U.S. Natl. Mus. Bull., 188: 1-622.
- Turner, B.J., 1965 A new place for the medakas. Classica, 1: (No.2) 1-6.
- Weber, M., 1913 Neue Beitrage zur Kenntnis der Süsswasserfische von Celebes. Bijd. Dierk., Amsterdam, 19: 197-213.
- Weber, M., and L.F. de Beaufort 1922 The fishes of the Indo-Australian archipelago IV. *Heteromii*, *Solenichthyes*, *Synentognathi*, *Percesoces*, *Labyrinthici*, *Microcyprini*. Leiden, E.J. Brill, Ltd.